Pt. Ravishankar Shukla University, Raipur

M. Sc. Biochemistry

Scheme and Syllabi of Examination for SESSION 2019-21

July 2019 - December 2019

	First Semester	Ma	rks	
Paper	Title of Paper	(External)	(Internal)*	Credit
1	Cell Biology	80	20	4
П	Biomolecules	80	20	4
Ш	Microbiology	80	20	4
IV	Biology of Immune System	80	20	4
LC-I	Lab Course I (Based on paper I & II)	80	20	2
LC-II	Lab Course II (Based on paper III & IV)	80	20	2
		Total	600	20

January 2020-June 2020

	Second Semester	Ma	rks	
Paper	Title of Paper	(External)	(Internal)*	Credit
1	Genetics and Molecular Biology	80	20	4
П	Bioenergetics & Metabolism	80	20	4
Ш	Instrumentation and Techniques	80	20	4
IV	Biometry, Computer and Scientometry	80	20	4
LC-I	Lab Course I (Based on paper I & II)	80	20	2
LC-II	Lab Course II (Based on paper III & IV)	80	20	2
		Total	600	20

July 2020-December 2020

	July 2020 2000			
	Third Semester	Ma	arks	
Paper	Title of Paper	(External)	(Internal)*	Credit
1	Genetic Engineering	80	20	4
41	Plant Physiology and Biochemistry	80	20	4
Ш	Environmental Biochemistry	80	20	4
IV	Enzymology	80	20	4
LC-I	Lab Course I (Based on paper I & II)	80	20	2
LC-II	Lab Course II (Based on paper III & IV)	80	20	2
		Total	600	20

January 2021-June 2021

	January 2022			
	Fourth Semester	Ma	arks	
Paper	Title of Paper	(External)	(Internal)*	Credit
I	Plant Biotechnology	80	20	4
Ш	Nutraceuticals and Functional Foods	80	20	4

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Ш	Special Paper-A: Clinical Biochemistry	80	20	4
	Special Paper-B: Molecular Endocrinology			
IV	Special Paper-A: Diagnostics & prophylaxis	80	20	4
	Special Paper-B: Bioinformatics			
LC-I	Lab Course I (Based on paper I & II)	80	20	2
LC-II	Lab Course I (Based on paper III & IV)	80	20	2
		Total	600	20

Important Note:

Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words.

Continuous evaluation of Performance*

Each student will be evaluated continuously throughout the semester. There will be a class test based on each theory paper. The full marks will be 10 for each paper. There will be a poster/oral presentation based on each theory paper. The full marks will be 10 for each presentation. Each student will be required to submit a brief write-up (not more than 15-20 pages) on his/her poster/oral presentation.

Project Work**

A student of IV semester will have the choice to opt for project work in lieu of four theory papers and two lab courses provided he/she secure at least 75% or more marks in aggregate in semester I and II. The project has to be carried out in recognized national laboratories or UGC-recognized universities. No student will be allowed to carry out project work in private laboratories/ college/ institutions, excluding the colleges recognized as research centers by the RDC of Pt. Ravishankar Shukla University, Raipur. The valuation of all the projects will be carried out by an external examiner and HoD of UTD or its nominee at the UTD Centre.

	r Lab Course (for each Semester)	Maximum Marks	100
External/I	nternal		
1-	Major exercise based on paper I	20	
2-	Minor exercise based on paper I	10	
3-	Major exercise based on paper II	20	
4-	Minor exercise based on paper II	10	
5-	Spotting/Interpretation*	10	
6-	Viva-voce	10	
Internal			
1-	Sessional	20	
Total		100	

A student will be required to interpret on the displayed item/material

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M. Sc. Biochemistry

FIRST SEMESTER (July 2019 – December 2019)

PAPER - I: CELL BIOLOGY

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- **UNIT-I Molecular organization of membranes** Asymmetrical organization of lipids, proteins and carbohydrates. Membrane transport: Passive transport, Osmosis, ion channels, membrane pumps and, Active transport: ATP-powered pumps-types, properties and mechanisms, electrical properties of membranes.
- **UNIT-II Protein trafficking:** Transport of proteins into mitochondria, chloroplast, endoplasmic reticulum and nucleus [in and out]. Transport by vesicle formation: exocytosis, endocytosis and its molecular mechanism.
- **UNIT-III Cell signaling**: Signaling via G-protein linked and enzyme linked cell surface receptors, MAP kinase pathways.

Eukaryotic cell division cycle: different phases and molecular events, regulation and control of cell cycle. Oncogenes: retinoblastoma, E2F and p53 proteins.

Apoptosis: regulation by CASPases and formation of apoptosome. Pro- and anti-apoptotic factors.

UNIT-IV States of chromosomes during cell cycle. Mitotic chromosome. Organization of genes in chromosomes. Banding pattern of chromosomes. Lampbrush and Polytene chromosomes. DNA packaging: Chromatin, nucleosomes, heterochromatin and euchromatin.

Lab Course:

- 1. Study of chromosome behaviour during Mitosis and meiosis (Onion / Garlic root tips, Onion buds, human lymphocytes, rat or bird testis /grass hopper testis or any other materials).
- 2. Calculation of mitotic index in growing Onion / Garlic root tips
- 3. Squash preparation: Polytene chromosome (in chironomus / Drosophila or other insect salivary gland) and Barr body (in buccal epithelial cells).
- 4. Demonstration of secretory granules in the salivary gland cells of insect.
- 5. Demonstration of mitochondria by vital staining.
- 6. Study of permanent slides.
- 7. Estimation of DNA
- 8. Estimation of RNA
- 9. Sub-cellular fractionation and marker enzymes
- 10. Identification of biomolecules in different tissues by histochemical techniques
- 11. Preparation of mitotic plate by carmine squashing method and phase identification.
- 12. Demonstration of the nuclear matrix networks in onion cells.
- 13. Study of the effect of chemical agents on chromosomes plant cells.
- 14. Isolation of protoplast, measurement of cell density plating efficiency.
- 15. Preparation of Karyotype of metaphase plate.
- 16. Preparation of Meiotic plate and determination of phases.

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- 17. Computation of Chaisma frequency and Terminalization of phases.
- 18. Micrometry and Camera Lucida drawings.

H. Lodish, A. Berk, S L Zipursky, P. Matsudaira D. Baltimore, and James Darnell.

B. Alberts, D. Bray, K. Hopkin, A. Johnson

H. Lodish, A. Berk, C. A. Kaiser & M. Krieger

B. Alberts, A. Johnson, J. Lewis and M. Raff Gerald Karp Molecular Cell Biology

Essential of Cell Biology Molecular cell Biology Molecular Biology of the Cell Cell and Molecular Biology Concepts and experiments

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FIRST SEMESTER (July 2019 – December 2019)

PAPER - II: Biomolecules

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I Classification, structure and function of Carbohydrates, Lipid:

Carbohydrate: Monosaccharides, homo and hetero-polysaccharides, Peptidoglycan glycoproteins and liposaccharide.

Lipids: Simple; cholestrol and complex; phospholipids and TAG

UNIT-II Classification, structure and function of amino acids, Synthesis of peptides, Proteins-properties, secondary, tertiary and quaternary structure of proteins, Ramchandran plot.
Nucleic Acid: Structure and function of Purine and pyrimidine, DNA-types, linking number, RNA-types.

UNIT-III Enzyme: apoenzymes, cofactors, coenzymes, active site, factors contributing to the catalytic efficiency of enzyme; enzyme kinetics- Michaelis-Menten equation, determination of Km, enzyme inhibition, allosteric enzymes, isoenzymes, multienzyme complexes

UNIT-IV Structure and biological role of: Porphyrins in biology, structure of hemoglobin and chlorophyll Animal hormones: protein, peptide and steroid hormones.

Vitamins: fat and water soluble.

Lab Course:

- 1. Specific tests for sugars, amino acids and lipids
- 2. Formal titration of amino acids
- 3. Estimation of proteins using ninhydrin and biuret method
- 4. Estimation of sugar by anthrone and Folin-Wu method.
- 5. Saponification value and iodine number of fat.
- 6. Estimation of ascorbic acid.
- 7. Achromic point determination using salivary amylase
- 8. Effect of ions on salivary amylase activity.
- 9. Enzyme assay and kinetics (ex. Amylase, Protease)

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Nelson, Cox and Lehninger

G. Zubay

Stryer

Garrett and Grosham

West, Tood, Mason & Bbruglen

White, Handler & Smith

D. Voet and J C Voet

Principles of Biochemistry

Biochemistry

Biochemistry

Biochemistry

Text book of biochemistry

Biochemistry-clinical application

Biochemistry

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FIRST SEMESTER (July 2019 – December 2019)

PAPER - III: Microbiology

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- UNIT-I General characteristics of fungi, classification of fungi, life cycle of selected fungal genus (Aspergillus, Pencillium, Fusarium and Mucor). Economic importance of fungi. Microbial association, parasitism, mutualism and symbiosis with plants and animals. Mycorrhiza, VAM. Algae: Distribution, classification, reproduction, ecology and importance.
- UNIT-II Morphology and ultra-structure of bacteria: Morphological types, cell wall of archaebacteria, gram negative, gram positive eubacteria. Bacterial cell membranes structure, composition and properties. Structure and function of flagella, cilia, pili, gas vesicles. Cyanobacteria, protozoa, mycoplasma and Rickettsia. Gene transfer mechanisms: transformation, transduction, conjugation and transfection. Plasmids and cosmid vector for gene cloning
- UNIT-III Nutritional types (autotrophs, heterotrophs, phototrophs, chemotrophs), growth curves, measurement of growth, factors affecting growth, generation time, growth kinetics. Batch and continuous culture, Basis of microbial classification, classification and salient feature of bacteria according to Bergey's manual of determinative bacteriology.
- UNIT-IV Viruses: Structure and classification; General concepts: Viral genome, capsids, envelopes, viroids and prions). Virus reproductions: Lysogeny and Lytic phase, Bacteriophages and their types. Introduction to Plant and animal viruses (TMV, HIV, Hepatitis virus, H1N1 virus, Small Pox virus and Ebola virus), Route of transmission of viruses, Laboratory diagnosis and treatment, Antiviral therapy

Lab Course:

- 1. Glassware preparation and sterilization techniques- wet heat- dry heat- filter types- laminar flow chamber types- CDC- safety levels.
- 2. Preparation of liquid & solid media, plating, pouring, inoculation and incubation for growth of microorganism
- Methods of obtaining pure culture of microorganisms (a) streak plate (b) Pour plate, and (c) spread plate methods
- 4. Identification and Microscopic examination of the microorganisms.

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- 5. Motility of bacteria by hanging drop technique.
- 6. Bacterial DNA isolation from E-coli culture.
- 7. Grams' staining for Gram positive and Gram's negative Bacteria.
- 8. Study of bacterial growth by turbiditimetry/ spectrophotometry
- 9. Isolation and enumeration of microorganisms from soil by serial dilution agar plating method.
- 10. Enumeration of viruses by plaque assay technique.

Microbiology

General Microbiology Principles of Microbiology

Microbiology

General Virology

Introduction to Mycology

Principles of Virology: Molecular

Biology, Pathogenesis, and Control of Animal Viruses

L.M. Prescott, J.P. Harley and D.A. Klein

RY Stanier, J L Ingrahamana, ML Wheelis & P. R. Painter

R.M. Atlas

Peleczar, Chan & Krieg.

Luria, Darnell, Baltimore and Campell.

CJ Alexopoulos and CW Mims

S. J. Flint, V. R. Racaniello, L. W. Enquist,

V. R. Rancaniello, A. M. Skalka

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FIRST SEMESTER (July 2019 - December 2019)

PAPER – IV: Biology of Immune System

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I Innate immune mechanism and characteristics of adaptive immune response. Cells of immune system: Hematopoisis and differentiation, mononuclear cells and granulocytes, antigen presenting cells.

Primary and Secondary lymphoid organs and tissues.

Ontogeny and phylogeny of lymphocytes. Lymphocyte traffic.

UNIT-II Antigen receptor molecules: B-cell receptor complex, Immunoglobulin- structure, types and function. T-cell receptor complex. Major Histocompatibility Complex- types, structural organization, function and distribution. Transplantation and Rejection. Complement system.

UNIT-III Antigens: nature of antigens, factor affecting immunogenicity, Haptens and super antigens. Antigenic determinants. Recognition of antigens by T and B cell.

Antigen processing. Role of MHC molecules in antigen presentation and co-stimulatory signals. Antigen and antibody interaction.

UNIT-IV Cell mediated immune response. Cytokines and interleukins- structure and function. Immunity to infections. Hypersensitive reactions and their types.

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Immunodeficiency disorders. Autoimmunity and autoimmune disorder. Immunological tolerance.

Lab Course:

- 1. Identification of cells of immune system
- 2. Separation of mononuclear cells by Ficoll-Hypaque
- 3. Identification of Lymphocytes and their subsets
- 4. Lymphoid organs and their microscopic organization
- 5. Isolation and purification of Antigens
- 6. Purification of IgG from serum
- 7. Estimation of Levels of gamma globulins and A/G ratio in blood
- 8. Antigen antibody interaction

Books Recommended:

Kuby's Immunology

R.A. Goldsby, T. J Kindt and B. A. Osborne

Immunology- A short Course

E. Benjamini, R. Coico and G. Sunshine

Immunology

Roitt, Brostoff and Male

Fundamentals of Immunology

William Paul Tizard

Immunology Immunology

Abbas et al

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SECOND SEMESTER (January 2020 – June 2020)

PAPER – I: Genetics and Molecular Biology

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-1 Gene mapping methods: Linkage maps, tetrad analysis, mapping with molecular markers, mapping by using somatic cell hybrids, development of mapping population in plants, complementation analysis.

Mutation: Types, mutagens and detection.

Mutant types — lethal, conditional, biochemical, loss of function, gain-of-function, germinal verses somatic mutants, insertional mutagenesis.

UNIT-II DNA replication in eukaryotes and prokaryotes: enzymes involved, replication origin and replication fork, fidelity of replication, extrachromosomal replicons.
 DNA damage and repair mechanisms: Repair of Base-excision, Nucleotide excisions, Mismatch and Double Strand. p₅₃ and p₂₁.

UNIT-III RNA synthesis and processing: enzymes involved, formation of initiation complex, transcription activator and repressor, elongation, and termination, RNA processing, capping, RNA editing, splicing, and polyadenylation, RNA transport.

UNIT-IV Protein synthesis and processing: Ribosome, formation of initiation complex, initiation factors, elongation and elongation factors and their regulation, termination.

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Aminoacylation of tRNA, tRNA-identity, aminoacyl tRNA synthetase, and translational proof-reading, translational inhibitors.

Post Translational modification of proteins.

Lab Course:

- 1. Isolation, purification and estimation of RNA
- 2. Isolation, purification and estimation of DNA
- 3. Determination of Tm of nucleic acid
- 4. Fraction of poly (A) RNA
- 5. Restriction Mapping
- 6. Restriction Digestion
- 7. Ligation
- 8. DNA molecular size determination

Books Recommended:

Molecular Cell Biology

H. Lodish, A. Berk, SL Zipursky, P. Matsudaira, D. Baltimore, and

James Darnell.

Essential Cell Biology

B. Alberts, D. Bray, K. Hopkin and A. Johnson

Molecular Biology of the Cell

B. Alberts, A. Johnson, J. Lewis and M. Raff

Cell and Molecular Biology : Concepts and experiments

Gerald Karp

Molecular Biology of the Gene

JD Watson et al.

Molecular Biology of the Cell

John Wilson, Tim Hunt

The Problems

John Wilson, Tillin

Molecular Biology of the Cell

Bruce Albert's, Alexander Johnson, Julian Lewis,

Martin Raff, Keith Roberts, Peter Walter

Genes VIII

Benjamin Lewin

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SECOND SEMESTER (January 2020 – June 2020)

PAPER - II: Bioenergetics & Metabolism

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I First and second laws of thermodynamics. Gibbs free energy G, free energy change ΔG , endergonic & exergonic reactions. Standard state free energy changes- ΔG , ΔG^0 and $\Delta G^{'0}$, Relationship between equilibrium constant and $\Delta G^{'0}$, Feasibility of reactions. ATP-Structure, properties and energy currency of the cell, Importance of Coupled reactions, other high energy compounds.

UNIT-II Carbohydrate metabolism: Glycolysis, Kreb's cycle, glycogenolysis, glycogenesis, pentose phosphate pathway, gluconeogenesis, and glyoxylate pathway. Regulation of carbohydrate metabolism.

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UNIT-III Electron transport and oxidation phosphorylation: electron carriers, complexes I to IV, substrate level phosphorylation, mechanism of oxidative phosphorylation. Shuttle system for entry of electron.

Biosynthesis and degradation of Lipids. Regulation of lipid metabolism

UNIT-IV Nitrogen Assimilation: Overview of Nitrogen in biosphere and uptake by organism. Biosynthesis and degradation of amino acids. Regulation of amino acid metabolism Biosynthesis and degradation of purine and pyrimidine nucleotides.

Lab Course:

- 1. Protein estimation by Lowry, Bradford and Spectrophotometric method
- 2. Estimation blood cholesterol
- 3. Estimation of sugar by Nelson-Somagy and Benedict's reagent
- 4. Isolation and estimation of lipid from seeds and egg.
- 5. Estimation of inorganic and total phosphorus by Fiske-Subba Rao method
- 6. Assay of phosphatases in blood and seeds
- 7. Urease estimation in plant tissues

Books Recommended:

Principles of Biochemistry

Biochemistry Biochemistry

Biochemistry

Text book of biochemistry

Biochemistry

Biochemistry with clinical application

Enzymes

Fundamentals of Enzymology

Practical biochemistry
Enzyme biotechnology
Enzyme Reaction Mechanism

Enzyme catalysis and regulation

Nelson, Cox and Lehninger

G. Zubay

Strver

Garrett and Grosham

West, Tood, Mason & Bbruglen

White, Handler & Smith

D. Voet and J C Voet

Dixon and Webb

Price and Steven

Plummer

G. Tripathi

Walsh

Hammes

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SECOND SEMESTER (January 2020 – June 2020)

PAPER- III: Instrumentation and Techniques

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I Centrifugation: Principle, techniques. Preparative, analytical and ultracentrifuges, sedimentation coefficient and factors affecting sedimentation coefficient. Application of centrifugation.

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Photometry: Basic principles of colorimetry, UV- visible spectrophotometry & IR-spectrophotometry. Spectroflurometry
Atomic absorption spectroscopy: Principle, Instrumentation and applications

UNIT-II Microscopic techniques: light microscopy, phase-contrast microscopy, scanning and transmission electron microscopy, different fixation and staining techniques for EM, freeze-etch and freeze-fracture methods for EM, image processing methods in microscopy.

UNIT-III Chromatography: Paper and Thin Layer Chromatography. Gel filtration, Ion exchange and Affinity chromatography. GLC and HPLC.
Histochemical and immunohistotechniques: Detection of molecules using immunoprecipitation, EIA, RIA and FIA.

UNIT-IV Electrophoresis: Agarose, PAGE, 2D-E.

Radioactivity: GM counter, liquid Scintillation counter, solid Scintillation counter, gamma

counters.

Lyophilization: Principle, instrumentation and applications.

Microtomy: types, principle and applications

Lab Course:

- Verification of Beers Law
- Determination of absorption maxima
- Quantitative determination, Enzyme kinetics
- Amino acid and carbohydrate separation by paper and TLC
- Ion exchange and gel filtration chromatography
- SDS Polyacralamide Gel Electrophoresis
- Isoenzymes
- Separation of sub-cellular organelles by differential centrifugation.
- Isolation of DNA and Agarose gel Electrophoresis

Books Recommended:

K Wilson and John Walker P

RF Boyer

Practical Biochemistry: Principles & Techniques Biochemistry Laboratory: Modern Theory &

Techniques

S Carson, H Miller and D Scott

Molecular Biology Techniques: A Classroom

Laboratory Manual

TC Ford and J. M. Graham

R Baserga and D Malamud

T Chard

Mark F. Vitha AGE Pearse An Introduction to Centrifugation

Autoradiography: techniques and application
An Introduction to Radioimmunoassay and Related

Techniques , Volume 6

TA Jennings Lyophilization: Introduction and Basic Principles
James M. Miller Chromatography: Concepts and Contrasts

Practical HPLC Method Development, 2nd Edition

A Handbook of Chromatography

Chromatography: Principles and Instrumentation

Histology and Histochemical methods

The principles of microscopy

Fundamentals of Light Microscopy and Electronic

Imaging, Second Edition

PA Midgley
DB Murphy & MW Davidson

LR Synder, JJ Kirkland and JL Glajch Anna Pratima Nikalje & D. Bhosale

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IB Buchwalow & W Bocker JB Birks

The Principles and Practice of Electron Microscopy Physical Principles of Electron Microscopy An Introduction to TEM, SEM, and AEM Freeze-Drying, 3rd Edition Practical Section Cutting and Staining Microtomy: Microtome and its applications

Immunohistochemistry and Immunocytochemistry: Essential Methods, Second Edition Immunohistochemistry: Basics and Methods The Theory and Practice of Scintillation Counting

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SECOND SEMESTER (January 2020 – June 2020)

PAPER- IV: BIOMETRY, COMPUTER AND SCIENTOMETRY

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- Unit-I Introduction to biostatistics. Types of biological data: data on different scales.
 Frequency distributions. Cumulative frequency distributions. Random sampling.
 Parameters and statistics. Measures of central tendency and dispersion: Mean,
 Median, Mode, Range, Variance and Standard deviation. Coefficient of variation.
 The effects of coding data. Data transformations: Log-transformation, Square-root transformation and Arcsine transformation. Distribution: normal & binomial.
 Probability: Basic laws of probability, addition law, multiplication law. Probability and frequency.
- Unit-II Statistical errors in hypothesis testing. Testing goodness of fit: Chi-square goodness of fit. Heterogeneity Chi-square. The 2 x 2 contingency table. One sample hypothesis. Two-sample hypothesis. Testing for difference between two means (t-test). Testing for difference between two variances (F-test). The paired sample t-test. Multiple-sample hypothesis (ANOVA): Single factor and two factors ANOVA. Multiple comparisons: Duncan's multiple-range tests. Simple linear regression. Regression vs. Correlation. Regression equation. Interpretations of regression functions. Simple linear correlation. The correlation coefficient.
- Unit-III Introduction to MS-Office software: Word processing; creating new document, editing documents, adding graphics to documents, Word tables. Management of Workbook & Worksheets; Applications, Features, Using formulas and functions, Features for Statistical data analysis, Excel ToolPak for data analysis, Generating charts/ graph. Presentation software; Working in PowerPoint, Creating new presentation, working with slides.

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Unit-IV Introduction to Internet and Applications. Basics of internet, e-mailing, Search engine – Google and Yahoo; Pub med, Scopus, Web of Science, Google Scholar, Indian Citation Index, Science Citation Index (SCI), h-index, i-10-index. Journal Impact Factor (JIF). Introduction to Plagiarism and Cyber laws.

Lab Course:

- 1. Exercises for data distribution
- 2. Exercises for computation of measures of central tendency
- 3. Exercises for computation of measures of variability
- 4. Computation of correlation coefficient, r, and regression constants
- 5. Data analysis by ANOVA and multiple-range tests
- 6. Hypothesis testing by t-test, F-test, and Chi-square test
- 7. Graphical presentation of data using a suitable package
- 8. Statistical analysis of a data using a suitable package
- 9. Preparation of document using a suitable package
- 10. Preparation of slides using a suitable package
- 11. Hands-on-practice for finding indices [SCI, h-index, i-10 index] of articles using relevant database

Books Recommended:

Campbell RC

Statistics for biologists

Zar JH

Biostatistical Analysis

Wardlaw AC

Practical Statistics for Experimental Biologists

Snedecor GW & Cochran WG

Statistical Methods

Sokal RR & Rohlf FJ

Introduction to Biostatistics

Computers: Concepts & Uses

Sumner M White R

How Computers Work

Cassel P et al.

Inside Microsoft Office Professional

Coleman P and Dyson P

Mastering Internets

Gralla P

How the Internet Works

Shelly GB, Vermaat ME,

Microsoft 2007: Introductory Concepts & Techniques

Cashman TJ

Microsoft Office 2003 All in One

Microsoft Office 2010 In Depth

Gilmore B

Habraken J

Plagiarism: Why it happens, How to prevent it?

Buranen L & Roy AM

Perspectives on Plagiarism & Intellectual Property

in a Post-Modern World

Kumar Anupa P

Cyber Law

Sood V

Cyber Law Simplified

M. Sc. Biochemistry THIRD SEMESTER (July 2020 - December 2020) PAPER – I: Genetic Engineering

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have guestions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

Molecular tools and their applications: restriction enzymes, modification enzymes.

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Molecular techniques: gel electrophoresis, polymerase chain reaction, DNA sequencing, DNA microarray. Hybridization techniques: western blot, northern blot, southern blot.

Gene cloning vectors: plasmids, lambda phage as vector, M13 phage as vector, cosmids, UNIT-II artificial chromosomes (BAC, PAC, YAC).

Library: 1. Genomic library: genomic DNA library construction and screening methods. 2. cDNA library: cDNA library construction and screening.

Study of gene regulation: reporter assays.

Expression strategies for heterologous genes: vector engineering and codon optimization, host engineering, in vitro transcription and translation.

UNIT-III Processing of recombinant proteins: recombinant proteins purification, refolding, characterization and stabilization, Applications of recombinant proteins Protein engineering; Site directed mutagenesis, Random mutagenesis and applications, Gene knockout technique

UNIT-IV Molecular Larkers: Mitochondrial markers: Ribosomal RNA (rRNA) genes; 12S rDNA and 16S rDN; protein coding genes; cytochrome b (cyt b), , NADH dehydrogenase subunit 5, cytochrome oxidase [DNA barcoding]

Chloroplast Markers: psbA-trnH, matK-trnK, ycf3-trnS, matK, and ycf1 [DNA barcoding]. Nuclear markers: RFLP, RAPD, AFLP, SSR, STS (Sequence Tagged Strands), microsatellites, SCAR (Sequence characterized amplified regions), SNP, SSCP (Single strand conformational polymorphism), DArT.

Lab Course:

- 1. Bacterial culture and antibiotic selection media. Preparation of competent cells
- 2. Isolation of plasmid DNA.
- 3. Isolation of Lambda phage DNA.
- 4. Quantitation of nucleic acids.
- 5. Agarose gel electrophoresis and restriction mapping of DNA.
- 6. Construction of restriction map of plasmid DNA.
- 7. Cloning in plasmid/phagemid vectors.
- 8. Isolation of RNA.
- 9. Synthesis of cDNA.
- 10. RAPD analysis by PCR.

Books Recommended:

Genes VIII

An Introduction to Genetic Engineering

Principles of Gene Manipulation and Genomics SB Primrose and Richard

Gene Cloning and Manipulation

Genetic Engineering (Genetics and Evolution)

Introduction to Biotechnology &

Genetic Engineering

Genetic Engineering

Biotechnology & Genetic Engineering

DNA Microarrays & Gene Expression: from

Experiments to Data Analysis and Modeling

DNA Sequencing (Intro. to Biotechniques)

Plant transformation Technologies

Application of Plant Biotechnology: In vitro

Benjamin Lewin

DST Nicholl

CJ Howe

R Hodge

AJ Nair

A Kumar & N Garg

L Yount

P Baldi & G Wesley

L Alphey

CN Stewart, A Touraev, V Citovsky & T Tzfira

A Kumar and SK Sopory

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Propagation, Pant Transformation and Secondary Metabolite Production Genetic Transformation of Plants

Transgenic Plants: Methods & Protocols

JF Jackson & HF Linskens

L Pena

M. Sc. Biochemistry THIRD SEMESTER (July 2020 – December 2020) PAPER- II: Plant Physiology and Biochemistry

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- UNIT- I Sensory photobiology Structure, function and mechanisms of action of phytochromes, cryptochromes and phototropins; stomatal movement; photoperiodism and biological clocks.
 Organization of shoot and root apical meristem; shoot and root development
- UNIT-II Photosynthesis: Light absorption and energy conversion, photosystems I and II, ATP synthesis, Assimilation of carbon in C₃, C₄ and CAM pathways, Photorespiration
- **UNIT-III** Phytohormones: Structure, biosynthesis, molecular mechanisms of Auxin, Gibberellins, Cytokinin, Abscisic acid and Ethylene, Brassinosteroids.
- UNIT-IV Senescence and Programmed cell death: Senescence; Metabolism and regulation of pigment and nucleic acid, PGR regulation, SAG. PCD; Formation of TE and mobilization of cereal endosperm, Formation of aerenchyma. Signal transduction and PCD

Lab Course:

- Spectrophotometric determination of chlorophyll-a, chlorophyll-b and total chlorophyll in young, mature and senescent leaves.
- 2. Kinetin estimation by cucumber cotyledons expansion bioassay.
- 3. Auxin bioassay using wheat coleoptiles.
- 4. GA bioassay by inducing de-novo synthesis of Amylase in de-embryonated seeds of wheat.
- 5. Estimation of mono, di and total phenols in the young and aged leaves.
- 6. Estimation of Guaiacol peroxidase activity in fresh and aged seeds.
- 7. Determination of Superoxide dismutase levels in the healthy and deteriorated seeds.
- 8. Estimation of metal toxicity induced changes in the AOS levels in leaf tissues.
- 9. Determination of Nitrate reductase activity in leaf tissues.
- 10. Separation of isozymes of SOD and GPX.

Books Recommended:

Fosket DF

Plant Growth & Development

Fover CH

Photosynthesis

Bacon KE

Photosynthesis: Photobiochem. & Photobiophysics

Leopold AC & Kriedemann PE

Plant Growth & Development

Moore TC

Biochemistry & Physiology of Hormones

L Taiz & E Zeiger

Plant Physiology

BB Buchanan, W Gruissem &

Biochemistry and Molecular Biology of Plants

RL Jones

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bengh on 06/9/19 William Wallet

MB Wilkins JA Hopkins FB Salisburry & CW Ross Hans-Walter Heldt

Advanced Plant Physiology Introduction to Plant Physiology Plant Physiology Plant Biochemistry & Molecular Biology

M. Sc. Biochemistry THIRD SEMESTER (July 2020 – December 2020) PAPER- III: Environmental Biochemistry

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have guestions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- UNIT-I Environmental Pollution: Major types. Outdoor and indoor air pollution, structure, sources, health effects and control strategies; water pollution, soil contamination, noise pollution and electromagnetic radiations: Sources, health effects and control strategies.
- Definition and scope of toxicology: Eco-toxicology and its environmental significance. Toxicity: Types, Acute and chronic exposures; toxicants and their classification .Health effects of toxic substances, Dose Response relationships; synergism and antagonism, determination of ED₅₀, LD₅₀, factors influencing toxicity. Toxicity testing – Test control, genetic toxicity testing.
- UNIT III Biochemical basis of toxicity: Mechanisms of Toxicity, disturbance of excitable membrane function, altered calcium homeostasis. Covalent binding to cellular macromolecules and genotoxicity. Tissue specificity of toxicity. Xenobiotic metabolism: Phase I reaction - oxidation - reduction, hydrolysis and hydration. Phase II reaction - conjugation and methylation.
- UNIT- IV Pesticide toxicity: Insecticides, fungicides, herbicides and biopesticides; Toxicology of food additives; Metal toxicity: arsenic, mercury, lead, cadmium and fluoride. Occupational hazards and risk assessment.

Lab Course:

- 1. Estimation of toxicants like fluoride and heavy metals in air, water and food.
- 2. Estimation of fluoride in human urine
- 3. Exposure of fishes to various concentrations of toxicants and assessment of their toxic effects by
 - a. Determination of percent mortality, LC 50 and LD50.
 - b. Determination of Bioconcentration Factor (BCF) of toxicants
 - c. Effect of various toxicants on serum enzymes, such as ASAT, ALAT, GPX, SOD, CAT and proteins
 - d. Demonstration of genotoxic effects of toxicants Comet Assay and Micronuclease assay

Books Recommended:

LG Corkerhem & BSS Shane DA Valler

Basic Environmental Toxicology Environmental Contaminants: Assessment and Control

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SVS Rana Ming Ho Yu

DW Sparling

Environmental Pollution: Health and Toxicology

Environmental Toxicology: Biological and health effects of

pollutants

Basics of Ecotoxicology

M. Sc. Biochemistry THIRD SEMESTER (July 2020 – December 2020) PAPER - IV: Enzymology

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I General properties and effects of pH, substrate and temperature on enzyme catalyzed reactions.

Kinetics of catalyzed reaction: Single substrate reactions, bisubstrate reactions, concept of Michaelis - Menten, Briggs Haldane relationship, Determination and significance of kinetic constants, Limitations of Michaelis-Menten Kinetics,

Concept of convergent and divergent evolution of enzyme.

UNIT-II Enzyme Turnover and methods employed to measure turnover of enzymes, significance of enzyme turnover.

Multienzyme system : occurrence, their properties , mechanism of action & regulation; Pyruvate dehydrogenase complex, fatty acid synthetase complexes.

Mechanism of action of lysozyme, chymotrypsin, carboxypeptidase and DNA polymease

UNIT-III General mechanisms of enzyme regulation

Allosteric enzymes, sigmoidal kinetics and their physiological significance, symmetrical and sequential modes for action of allosteric enzymes and their significance.

Water soluble enzymes and their coenzymes. Metallo enzymes.

Immobilized enzymes and their industrial applications.

UNIT-IV Enzymes of Industrial Importance; their source, characteristic properties,

functions and uses.

Enzymes used in leather, paper, textile industries.

Enzymes in baking, brewing, Alcohol products; enzymes in detergents, starch and animal feeds.

Amylases, cellulases, catalase, pectinase, lipase, protease, xylanase, laccase, beta glucanase

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Lab Course:

- 1. Estimation of enzymes
- 2. Separation, purification of sub-cellular organelles & assay of marker enzymes.
- 3. Methods of purification of an enzyme ion-exchange, gel filtration
- 4. Test of homogeneity by SDS-PAG E
- 5. Kinetics of an enzymatic reaction
- 6. Effect of various toxicants on serum enzymes and proteins
- 7. Enzyme modeling: Validation Criteria by WHATIF, Verify3d, PROSA and DOPE score
- 8. Verification of Ramachandran Plot: Estimation of interaction energy per residue by PROSA and Verify3D.
- 10. Enzyme packing quality: Assessed by WHATIF.

Books Recommended:

Brandon and Tooze

Introduction to Protein Structure

Campell

Discovering Genomics, Proteomics and Bioinformatics,

Dan Gusfield

Algorithms on Strings Trees and Sequences

Lesk, A.M

Introduction to Protein Architecture

Mcpherson, A.

Introduction of Molecular Crystallography

Pennington

Proteomics from Protein Sequence to Function

Durbin, Eddy, Anders & Graeme

Biological Seq. Analysis: Probabilistic Models of Proteins &

Nucleic Acids

S.A. Bbernhard

The structure and function of enzymes

J. Palmer

Enzymes: biochemistry, Biotechnology, Clinical chemistry

M Dixon, EC Webb, CJR Thorne

Enzymes

& KF Tipton

Alan Fersht

Enzyme structure and Mechanism Enzymatic reaction mechanism

Christopher Walsh Eisenthal and Danson

Enzyme Assay: A Practical Approach

M. Stipanuk

Biochemical, Phys. & Mol. Aspects of Human Nutrition

G.P. Talwar

Text book of Biochemistry and Human Biology

M. Sc. Biochemistry FOURTH SEMESTER (July 2020 – December 2020) PAPER – I: Plant Biotechnology

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT- I Introduction to cell and tissue culture: Tissue culture media (composition and preparation), explant preparation, Callus and suspension culture, cytodifferentiation and organogenic differentiation, somatic embryogenesis, micropropagation.

Shoot tip culture: Rapid clonal propagation and production of virus free plant.

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Pendica 06/5/19 UNIT-II Embryo culture and embryo rescue.

Anther, pollen and ovary culture for production of haploid plants & homozygous lines.

Protoplast isolation, culture and fusion; selection of hybrid cells and regeneration of hybrid plants; symmetric and asymmetric hybrids, cybrids.

Germplasm conservation: Cryopreservation & slow growth cultures.

Chloroplast Transformation: Advantages, vectors, success; tobacco & potato.

UNIT-III Plant transformation technology: Plant transformation technology: basis of tumor formation, hairy root, features of Ti and Ri plasmids, mechanism of DNA transfer, role of virulence genes, use Ti and Ri as vectors, binary vectors, use of 35S and other promoters, use of reporter genes, particle bombardment, electroporation, microinjection.

UNIT-IV Applications of plant transformation for productivity and performance: herbicide resistance, insect resistance, long shelf-life of fruits. Bt genes, Non–Bt like protease inhibitors & amylase inhibitors, virus resistance, nucleocapsid gene, disease resistance, PR (Pathogenesis Related) proteins, nematode resistance, abiotic stress, male sterile lines

Lab Course:

- 1. Preparation of culture media.
- 2. To performe meristem/ bud culture, shoot multiplication & rooting phenomenon.
- 3. To study organogenesis.
- 4. To perform somatic embryogenesis.
- 5. To study the process of plantlet acclimatization.
- 6. To perform embryo culture.
- 7. To study the process of anther culture development.
- 8. Study of molecular markers.
- 9. Extraction of DNA from plant cultures.
- 10. Estimation & separation of DNA: Agarose gel electrophoresis & spectrophotometer.

Books Recommended:

Razdan MK Introduction to Plant Tissue Culture

Vasil IK Plant Cell and Tissue Culture
Bhojwani SS and Razdan MK Plant Tissue Culture

Fu TJ, Singh G and Curtis WR Plant Cell & Tissue Culture for the production of Food Ingredients

Hammond, McGarvP & Yusibov Plant Biotechnology

Singh BD Biotechnology: Expanding Horizons

RH Smith Plant Tissue Culture Techniques and Experiments
L Kyte and J Kleyn Plants from Test Tubes: An Introduction to Micropropagation

M Smith Plant Propagator's Bible

MR Ahuja Micropropagation of Woody Plants
YPS Bajaj Trees III

YPS Bajaj Trees IV

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M. Sc. Biochemistry

FOURTH SEMESTER (January 2021 – June 2021)

PAPER- II: Nutraceutical and Functional Foods

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

Unit-I: Introduction to Nutraceuticals as Science:

Historical perspective, classification, scope and future prospects. Scrutinising the term 'nutraceutical', Regulation of various countries. Medicinal Plants: Ethnomedicine in India, Applied aspects of the Nutraceutical Science. Sources of Nutraceuticals. Relation of Nutraceutical Science with other Sciences: Medicine, Human physiology, genetics, food technology, chemistry and nutrition

Unit-II: Properties, structure and functions of various Nutraceuticals:

Glucosamine, Octacosanol, Lycopene, Falvanoids, Carnitine, Melatonin and Ornithine alpha, ketoglutarate. Use of proanthocyanidins, grape products, flaxseed oil as Nutraceuticals. Nutraceutical Industry and Market Information, New technologies in development of Nutraceuticals and functional foods

Functional Foods, Scope of Genetic engineering, Nutritional Genomics

Unit-III: Food as remedies

Nutraceuticals bridging the gap between food and drug, Special Dietary Needs, Disease and Nutrition; Nutraceuticals in treatment for cognitive decline, Nutraceutical remedies for common disorders like Arthritis, Bronchitis, circulatory problems, hypoglycemia, Nephrological disorders, Liver disorders, Osteoporosis,

Psoriasis and Ulcers etc. Brief idea about some Nutraceutical rich supplements e.g. Bee pollen, Caffeine, Green tea, Lecithin, Mushroom extract, Chlorophyll, Kelp and Spirulina etc.

Unit-IV: Anti-nutritional Factors present in Foods

Types of inhibitors present in various foods and how they can be inactivated. General idea about role of Probiotics and Prebiotics as nutraceuticals. Recent advances in techniques & feeding of substrates. Assessment of nutritional status and Recommended Daily allowances.

Lab Course:

- Identification using characteristic features of nutracuetically important plants like; *Phyllanthus emblica, Curcuma longa, Zinziber officinalis*, Solanaceae (*Withania somnifera*), *Aloe vera*, Lilliaceae (*Alium sativum*), Lamiaceae (*Ocimum sanctum*), Apiaceae (*Coriandrum sps*) and Liliaceae (*Asparagus sps.*), *Centella asiatica*.
- Study of following Parasites/ Vectors/ pests: Identification, Habits and control measures (museum Specimens / slides): Entamoeba histolytica, Taenia sps, Ascaris lumbricoides, Ancylostoma dueodenaei, Trichinella spiralis, Trichura trichuris, Mosquito (Culex and Anopheles), House fly, Green bottle fly, Head Louse, Cockroach (Periplanata & Blatta), bed bug, Mus sps. (Mouse) and Rattus sps. (House rat)

Reactions of mono, di and polysaccharides and their identification in unknown mixtures

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- Determination of Acid value, Saponification and Iodine number of natural fats & oils.
- Estimation of proteins with Bradford's and other methods.
- Extraction and estimation of total sugars from food products (dairy product, fruit juices, bread).
- TLC separation of Plant pigments Curcumin and carotene.
- To isolate DNA and RNA from given plant/ animal material and estimate DNA by Diphenylamine (DPA) method and RNA by Orcinol reagent
- · Extraction, purification and evaluation of activity of any one digestive enzyme (e.g. Beta amylase from sweet potato)
- Estimation of ascorbic acid from lemon & amla juice by titration method
- Estimation of crude fat contents of foods by Soxhlet's method (Butter, Margarine, edible oil).
- Estimation of total Nitrogen of foods by Kjeldahl and Micro Kjeldahl methods.
- Chromatography: Paper, TLC, adsorption, ion exchange, gel filtration, affinity, GC & HPLC.
- Separation of Milk proteins on Native and SDS gels.
- Preparation of plasmid DNA from given sample and its digestion by restriction enzymes and separation of DNA fragments by gel electrophoresis

01. Stryer E.A.,	Biochemistry
02. Zubay, Geoffrey L.	Biochemistry,
03. Greenberg David M.	Metabolic Pathways, Vol 3
04. Todd and others,	Clinical Diagnosis and Management, 17th Ed,
05. Gopalan C., et al	Dietary Allowances for Indians, NIH, Hyderabad.
06. Anita F.P.	Clinical Dietetics and Nutrition, 4th Ed, 1997,
07. Devlin, T.M.	Text Book of Biochemistry with Clinical Correlation,
08. Mahan, L.K. & Ecott- Stump, S. [Ed.]	Krause's Food, Nutrition and Diet Therapy
09. Lehninger	Nutrition Concepts & Controversies,
10. Davidson, S. Passmore, & Turswell	Nutrition and dietetics
10. Goodhearth R., S. Shills	Modern Nutrition in health and disease
12. Nelson and Cox, 2000,	Lehninger's Principles of Biochemistry,
13. Robert E.C. Wildman,	Handbook of Nutraceuticals and Functional Foods
16. Rapport and B. Lockwood	Nutraceuticals
15. W. Jeffrey, Hursts	Methods of Analysis for Functional Foods and
	Nutraceuticals
16. M. Maffei (Ed.)	Dietary Supplements of Plant Origin
17. Gunzler and Williams	Handbook of Analytical Techniques Vol. I,II,
18. Thomson	Herbal Medicines PDR 3rd ed
19. Gary, M & Giintert, E.	Active Compounds in Foods Chemistry and Sensory
	Properties - I
20. Israel Goldberg	Functional foods, designer foods, pharma foods,
	Nutraceuticals,
21. P.D. Dabre	Introduction to Practical Molecular Biology,
22. Ellyn Daugherty	Biotechnology Science for the new Millennium,
23. T K Attwood, D J PSmith	Bioinformatics Introduction
24. Primrose and RM Twyman	Principals of Gene Manipulation and Genomics.
25. Massimo Maffei	Dietary Supplements of Plant origin: a nutrition and
	health approach
27. CC RUM	Herbal Drugs: Potential Antimalarial Herbal Drugs from
	South Asia.

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M. Sc. Biochemistry

FOURTH SEMESTER (January 2021 – June 2021)

Special Paper PAPER - III (A): Clinical Biochemistry

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- UNIT-I: Plasma proteins Properties, functions and their variations in diseases, Plasma lipids and lipoproteins, Interrelationship of lipids, lipoproteins and apolipoproteins. Erythropoiesis, abnormalities in blood formation. Anemias. Heamoglobinopathies. Cerebrospinal fluid composition in health and diseases. Plasma enzymes in diagnosis and prognosis.
- **UNIT-II:** Kidney & Liver function tests: Renal function tests, osmolarity and free water clearances, acute and chronic renal failure. Renal hormones—Renin, erythropoietin and angiotensin. Liver function tests: clinical features and test based on bile pigments level, prothrombin time.
- **UNIT-III:** Gastric function tests: collection of gastric contents, examination of gastric residium, Fractional Test Meal, stimulation tests, tubeless gastric analysis. Gastrointestinal hormones Gastrin, secretin and cholecystokinin. Disorders of gastric function. Pancreatic exocrine secretions and pancreatic diseases.
- UNIT-IV: Molecular diagnosis of genetic defects: Diagnosis of genetic diseases by molecular techniques (cystic fibrosis, Hemachromatosis, thalassemias, sickle diseases) DNA probes; restriction fragment length polymorphism (RFLP); polymerase chain reaction (PCR); amplification of mRNA. AIDS: Clinical diagnosis.

Lab Course:

- 1. Assay of Alkaline and Acid Phosphates
- 2. Estimation of blood glucose by GOD and POD method
- 3. Various types of glucose tolerance tests.
- 4. Estimation of SGOT, SGPT, LDH and CPK, Serum Amylase enzymes
- 5. Estimation of HDL- cholesterol, LDL- cholesterol.
- 6. Estimation of uric acid and creatinine in plasma.
- 7. Estimation of urine and blood billurubin.

Books Recommended:

Textbook of Biochemistry with Clinical Correlations: T.M. Devlin

Lippincott's Illustrated Reviews in Biochemistry: P.C.Champe, R.A.Harvey and D.R.Ferrier

Harper's Biochemistry : R.K.Murray, D.K.Granner, P.A. Mayes & V.W.Rodwell

Clinical Laboratory Science Review: Robert R. Harr

Fundamentals of Clinical Chemistry: C.A. Burtis, E.R. Ashwood Tietz

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Notes on Clinical Chemistry-: Whitby, Smith, Beckett, Walker, Harrison

The structure and function of enzymes: S.A. Bbernhard

Enzymes- biochemistry, Biotechnology, Clinical chemistry: J. Palmer

Enzymes: Dixon, Webb, Thorne & Tipton Enzyme structure and Mechanism: Alan Fersht Enzymatic reaction mechanism: C. Walsh, F. Pub

Enzyme Assay: A Practical Approach: Eisenthal and Danson

M. Sc. Biochemistry

FOURTH SEMESTER (January 2021 – June 2021)

Special Paper: PAPER- III (B): Molecular Endocrinology

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

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Unit-I	Definition and scope of molecular endocrinology. Chemical nature and general classes of hormones: Peptide, Amino acid derived, Steroid, Neurotransmitters, Neuropeptides, Chalones, Eicosanoids and Pheromones. Hypothalamic octapeptide hormones: Oxytocin and Vasopressin. Purification and characterization of hormones. Hypothalamohypophyseal axis.
Unit-II	Genetic control of hormone synthesis: Structure and expression of protein hormone encoding gene. Molecular aspects of peptide hormone biosynthesis and secretion. Molecular aspects of synthesis and delivery of thyroid hormones, biogenic amines and steroid hormones. Production of protein hormones by recombinant DNA technology
Unit-III	Molecular mechanism of hormone action: Membrane, cytoplasmic and nuclear hormone receptors, Non-genomic mechanism of hormone action, Receptor-ligand interactions. Hormonal signal transduction: G-proteins and second messengers. Genomic mechanism of hormone action: Steroid and thyroid hormones.
Unit-IV	Molecular aspects of Reproductive endocrinology: Genetics of sex. Testicular and ovarian determining genes. Mullerian inhibiting substance genes. Stem cell renewal in testis. Molecular basis of male and female contraception. Endocrine disruptors. Neuroendocrine control of reproduction and feedback mechanism.
Lab Course	
1.	Purification of any protein hormone.
2.	Assay of steroid dehydrogenase
3.	Isolation and characterization of steroid/prostaglandin.

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4.	In vivo bio- assay for estrogen/testosterone/LH
5.	Perfusion technique for the fixation of endocrine tissue
6.	Identification of hypothalamic nuclei following histological / histochemical methods
7.	Histological / Cytological / Histochemical study of endocrine glands.
8.	Study of estrus cycle by vaginal smear technique.
9.	Extraction and estimation of Gondotrophin / Pregnanediole from urine.
10.	Sperm count and motility.
11.	Study of neurosecretory cells/ materials/granules in the given materials.

Recommended Books

Franklyn F. Bolander Freedman and Birkhauser

Brown R.
Mac E. Hadley
D Groot. L. J. (ed.), W. B. Saunder
Norris, D. O.
Brook, C.G.D. and Marshall, N.J.
Shlomo Melmed et al
Goodman. H.M.

Negi. Yen et al (ed) Adashi et al, Zarrow et al.

Chinov et al.

Claude D and Larry L. E (ed)

Litwack, G.

CM Bunce, MJ Campbell

Molecular Endocrinology

Molecular Biology of Steroid and Nuclear Hormone

receptors:

An Introduction to Neuroendocrinology

Endocrinology

Endocrinology (Vol. I-III) Vertebrate Endocrinology Essential Endocrinology

Williams Textbook of Endocrinology

Basic Medical Endocrinology Introduction to Endocrinology Reproductive Endocrinology Reproductive Endocrinology

Experimental Endocrinology
Essential techniques in reproductive
physiology and Endocrinology

Cell and Molecular Biology of Testis Biochemical actions of hormones Nuclear Receptors: Current Concepts

and Future Challenges

M. Sc. Biochemistry

FOURTH SEMESTER (January 2021 – June 2021)

Special Paper: PAPER- IV (A): Diagnostics and prophylaxis

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-1 Generation of diversity in BCR. Light and heavy chain gene recombination. Recombination Signal sequences. Class switching. Membrane and secreted immunoglobulins. Organization, arrangement of T-cell receptor genes and recombination.

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Regulation of immune response by antigen, antibody, immune complex, MHC and cytokines.

- UNIT-II Immunoprophylaxis: Principles of Vaccination. Immunization practices. Vaccines against important bacterial and parasitic diseases. DNA vaccines; passive prophylactic measures. Viral vaccines and antiviral agents. Vaccination schedules and safety. Production of vaccines.
- UNIT-III Diagnosis of microbial diseases Collection, transport and preliminary processing of Clinical pathogens. Clinical, microbiological, immunological and molecular diagnosis of diseases. Principles of immunodiagnostics. Antigen-antibody based diagnosis and the techniques involved – Enzyme, Radio and Fluorescence Immuno assays, Immunoblotting, Flow cytometry. Effector cell assays, Cytotoxic assays. Isolation of pure antibody. Monoclonal & Designer antibody and their application in immunodiagnostics.
- UNIT-IV Modern methods of microbial diagnosis. Use of nanotechnology in diagnosis. Synthesis of Nanomaterials, Nanoparticle based drug delivery, Toxicity and environmental risks of nanomaterials.

Biosensors: Biosensor-development, types and characteristics, DNA biosensors, application of biosensors in clinical diagnostics: detection of infectious diseases, food pathogen and environmental monitoring.

Lab Course:

- 1. Preparation of Parasite/microbe Antigen and analysis by PAGE
- 2. Immunizations and Production of Antibody
- 3. Antigen antibody reaction by Double Diffusion, Counter Current and IEP, RID and ELISA
- 4. Western Blot Analysis
- 5. Immunodiagnosis using commercial kits (VDRL, RPR, Widal etc.)
- 6. Identifications of nanomaterials using physical and chemical properties.
- 7. Green and chemical route for synthesis of nanomaterials.
- 8. Nanomaterial characterizations using UV-Vis and FT-IR spectroscopy.
- 9. Assessment of antibacterial properties of nanomaterials.
- 10. Identification of different analyte/ biomolecules for biosensing system.

Books Recommended:

R.A. Goldsby, T.J Kindt & B. A. Osborne E. Benjamini, R. Coico and G. Sunshine

Roitt, Brostoff and Male

William Paul Stewart Snell

Elgert

M. Wilson, K. Kannangara, G Smith, Simmons, B. Raguse

G. Cao

Challa S.S.R. Kumar

Charles P. Poole Jr. and Franks. J. Qwens

C. M. Niemeyer, C. A. Mirkin (Editor)

Kuby's Immunology:

Immunology-A short Course

immunology

Fundamentals of Immunology

Immunology, Immunopathology and Immunity

Understanding Immune System

Basic science and Emerging M. Nanotechnology:

technologies,

Nanostructures and Nanomaterials: Synthesis, properties and

applications

Nanomaterials for medical diagnosis and therapy,

Introduction to Nanotechnology

Nanobiotechnology: Concepts, Applications and Perspectives

M. Sc. Biochemistry

FOURTH SEMESTER (January 2021 – June 2021)

Special Paper: PAPER- IV (B): Bioinformatics

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have guestions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

Introduction to bioinformatics and data generation Unit I

Bioinformatics and its relation with molecular biology. Examples of related tools (FASTA, BLAST, BLAT, RASMOL), databases (GENBANK, Pub med, PDB) and software (RASMOL, Ligand Explorer).

Data generation; Generation of large scale molecular biology data. (Through Genome sequencing, Protein sequencing, Gel electrophoresis, NMR Spectroscopy, X-Ray Diffraction, and microarray). Applications of Bioinformatics.

Unit II **Biological Database and its Types**

Introduction to data types and Source. Population and sample. Classification and Presentation of Data. Quality of data, private and public data sources. General Introduction of Biological Databases; Nucleic acid databases (NCBI, DDBJ, and EMBL). Protein databases (Primary, Composite, and Secondary). Specialized Genome databases: (SGD, TIGR, and ACeDB). Structure databases (CATH, SCOP, and PDB sum)

Unit III Data storage and retrieval and Interoperability

Flat files, relational, object oriented databases and controlled vocabularies. File Format (Genbank, DDBJ, FASTA, PDB, SwissProt). Introduction to Metadata and search; Indices, Boolean, Fuzzy, Neighboring search. Data exchange and integration. Ontologies, interchange languages and standardization efforts.

General Introduction to XML, UMLS, CORBA, PYTHON and OMG/LIFESCIENCE.

Unit IV Gene Expression and Representation of patterns and relationship

General introduction to Gene expression in prokaryotes and eukaryotes, transcription factors binding sites. SNP, EST, STS.

Regular Expression, Hierarchies, and Graphical models (including Marcov chain and Bayes notes). Genetic variability and connections to clinical data.

Lab Course:

- 01. Retrieval of sequences from NCBI, EBI and EMBL databases.
- 02. Retrieval of sequences from NBRF-PIR, SWISSPROT and P databases.
- 03. Transition and Translation of sequences.
- 04. Retrieval of genome from genome databases.
- 05. Exploring DIP and PPI.
- 06. Exploring BIND and PIM.
- 07. Exploring MINT and GRID.
- 08. Analysis of phylogenetic tree
- 09. Exploring PDB file.
- 10. Analysis of active site by pymol

BAXEVANIS, AD & OUELLETTE, BFF: Bioinformatics: a practical guide to the analysis of

genes and proteins. 2nd Ed.. 2002.

BAXEVANIS, AD, DAVISON, DB, PAGE: Current protocols in bioinformatics. 2004.

RDM & PETSKO, GA

ORENGO, C, JONES, D &

: Bioinformatics: genes, proteins and computers. 2003

THORNTON, J

Ingvar Eidhammer, Inge Jonassen, : Protein Bioinformatics. 2003

William R Taylor

HIGGINS, D & TAYLOR, W

: Bioinformatics: sequence, structure, and databank. 2000.

David Mount

: Bioinformatics: sequence and genome analysis. 2004