

Pt. Ravishankar Shukla University, Raipur

M. Sc. BIOCHEMISTRY

Scheme and Syllabi of Examination for SESSION 2018-20

July 2018-December 2018				
	First Semester	Marks		Credit
Paper	Title of Paper	(External)	(Internal)*	
I	Cell Biology	80	20	4
II	Biomolecules	80	20	4
III	Microbiology	80	20	4
IV	Biology of Immune System	80	20	4
LC-I	Lab Course I (Based on paper I & II)	80	20	2
LC-II	Lab Course II (Based on paper III & IV)	80	20	2
	Total	600	20	
January 2019-June 2019				
	Second Semester	Marks		Credit
Paper	Title of Paper	(External)	(Internal)*	
I	Genetics and Molecular Biology	80	20	4
II	Bioenergetics & Metabolism	80	20	4
III	Instrumentation and Molecular Techniques	80	20	4
IV	Biometry, Computer and Scientometry	80	20	4
LC-I	Lab Course I (Based on paper I & II)	80	20	2
LC-II	Lab Course II (Based on paper III & IV)	80	20	2
	Total	600	20	
July 2019-December 2019				
	Third Semester	Marks		Credit
Paper No.	Title of Paper	(External)	(Internal)*	
I	Genetic Engineering	80	20	4
II	Plant Physiology and Biochemistry	80	20	4
III	Nutritional and Environmental Biochemistry	80	20	4
IV	Enzymology	80	20	4
LC-I	Lab Course I (Based on paper I & II)	80	20	2
LC-II	Lab Course II (Based on paper III & IV)	80	20	2
	Total	600	20	
January 2020-June 2020				
	Fourth Semester	Marks		Credit
Paper No.	Title of Paper	(External)	(Internal)*	
I	Plant Biotechnology	80	20	4
II	Seed Science Technology	80	20	4
III	Special Paper-A: Clinical Biochemistry and Endocrinology Special Paper-B: Nutraceuticals and Functional Foods	80	20	4
IV	Special Paper-A: Advanced Immunology, diagnostics and prophylaxis Special Paper-B: Bioinformatics	80	20	4
LC-I	Lab Course I (Based on paper I & II)	80	20	2
LC-II	Lab Course I (Based on paper III & IV)	80	20	2

		Total	600	20
	OR			
	Project Work**	Dissertation	240	60
		Seminar based on Projects	160	40
		Viva-voce	80	20
		Total	600	20
		Grand Total	2400	Credit: 80

Important Note:

Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words.

Continuous evaluation of Performance*

Each student will be evaluated continuously throughout the semester. There will be a class test based on each theory paper. The full marks will be 10 for each paper. There will be a poster/oral presentation based on each theory paper. The full marks will be 10 for each presentation. Each student will be required to submit a brief write-up (not more than 15-20 pages) on his/her poster/oral presentation.

Project Work**

A student of IV semester will have the choice to opt for project work in lieu of four theory papers and two lab courses provided he/she secure at least **75%** or more marks in aggregate in semester I and II. The project has to be carried out in recognized national laboratories or UGC-recognized universities. No student will be allowed to carry out project work in private laboratories/ college/ institutions, excluding the colleges recognized as research centers by the RDC of Pt. Ravishankar Shukla University, Raipur. The valuation of all the projects will be carried out by an external examiner and HoD of UTD or its nominee at the UTD Centre.

Scheme for Lab Course (for each Semester)

Maximum Marks 100

1- Major exercise based on paper I	20
2- Minor exercise based on paper I	10
3- Major exercise based on paper II	20
4- Minor exercise based on paper II	10
5- Spotting/ Interpretation*	10
6- Viva-voce	10
7- Sessional [Internal]	20
Total	100

* A student will be required to interpret on the displayed item/material

M. Sc. Biochemistry
FIRST SEMESTER (July 2018 – December 2018)
PAPER - I: CELL BIOLOGY [Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I Molecular organization of membranes - Asymmetrical organization of lipids, proteins and carbohydrates. Membrane transport: Passive transport, Osmosis, ion channels, membrane pumps and, Active transport: ATP-powered pumps-types, properties and mechanisms, electrical properties of membranes.

UNIT-II Protein trafficking: Transport of proteins into mitochondria, chloroplast, endoplasmic reticulum and nucleus. Transport by vesicle formation: exocytosis, endocytosis and its molecular mechanism.

UNIT-III Cell signaling: Signaling via G-protein linked and enzyme linked cell surface receptors, MAP kinase pathways.

Eukaryotic cell division cycle: different phases and molecular events, regulation and control of cell cycle. Oncogenes: retinoblastoma, E2F and p53 proteins.

Apoptosis: regulation by CASPases and formation of apoptosome. Pro- and anti-apoptotic factors.

UNIT-IV States of chromosomes during cell cycle. Mitotic chromosome. Organization of genes in chromosomes. Banding pattern of chromosomes. Lampbrush and Polytene chromosomes. DNA packaging: Chromatin, nucleosomes, heterochromatin and euchromatin.

Lab Course:

1. Study of chromosome behaviour during Mitosis and meiosis (Onion / Garlic root tips, Onion buds, human lymphocytes, rat or bird testis /grass hopper testis or any other materials).
2. Calculation of mitotic index in growing Onion / Garlic root tips
3. Squash preparation: Polytene chromosome (in chironomus / Drosophila or other insect salivary gland) and Barr body (in buccal epithelial cells).
4. Demonstration of secretory granules in the salivary gland cells of insect.
5. Demonstration of mitochondria by vital staining.
6. Study of permanent slides.
7. Estimation of DNA
8. Estimation of RNA
9. Sub-cellular fractionation and marker enzymes
10. Identification of biomolecules in different tissues by histochemical techniques

11. Preparation of mitotic plate by carmine squashing method and phase identification.
12. Demonstration of the nuclear matrix networks in onion cells.
13. Study of the effect of chemical agents on chromosomes plant cells.
14. Isolation of protoplast, measurement of cell density plating efficiency.
15. Preparation of Karyotype of metaphase plate.
16. Preparation of Meiotic plate and determination of phases.
17. Computation of Chiasma frequency and Terminalization of phases.
18. Micrometry and Camera Lucida drawings.

Books Recommended:

- | | |
|---|-------------------------------|
| H. Lodish, A. Berk, S L Zipursky, P. Matsudaira | Molecular Cell Biology |
| D. Baltimore, and James Darnell. | |
| B. Alberts, D. Bray, K. Hopkin, A. Johnson | Essential of Cell Biology |
| H. Lodish, A. Berk, C. A. Kaiser & M. Krieger | Molecular cell Biology |
| B. Alberts, A. Johnson, J. Lewis and M. Raff | Molecular Biology of the Cell |
| Gerald Karp | Cell and Molecular Biology |
| | Concepts and experiments |

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FIRST SEMESTER (July 2018 – December 2018)

PAPER – II: Biomolecules [Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I Classification and function of Carbohydrates, Lipid:

Carbohydrate: Monosaccharides, homo and hetero-polysaccharides, Peptidoglycan glycoproteins and liposaccharide.

Lipids: Simple; cholesterol and complex; phospholipids and TAG

UNIT-II Classification and functions of amino acids, Synthesis of peptides, Proteins- properties, secondary, tertiary and quaternary structure of proteins, Ramchandran plot.

Nucleic Acid: Purine and pyrimidine, DNA-types, linking number, RNA-types.

UNIT-III Enzyme: apoenzymes, cofactors, coenzymes, active site, factors contributing to the catalytic efficiency of enzyme; enzyme kinetics- Michaelis-Menten equation, determination of Km, enzyme inhibition, allosteric enzymes, isoenzymes, multienzyme complexes

UNIT-IV Structure and biological role of:

Porphyrins in biology, structure of hemoglobin and chlorophyll

Animal hormones: protein, peptide and steroid hormones.

Vitamins: fat and water soluble.

Lab Course:

1. Specific tests for sugars, amino acids and lipids

2. Formal titration of amino acids
3. Estimation of proteins using ninhydrin and biuret method
4. Estimation of sugar by anthrone and Folin-Wu method.
5. Saponification value and iodine number of fat.
6. Estimation of ascorbic acid.
7. Achromic point determination using salivary amylase
8. Effect of ions on salivary amylase activity.
9. Enzyme assay and kinetics (ex. Amylase, Protease)

Books Recommended:

Nelson, Cox and Lehninger	Principles of Biochemistry
G. Zubay	Biochemistry
Stryer	Biochemistry
Garrett and Grosham	Biochemistry
West, Tood, Mason & Bbruglen	Text book of biochemistry
White, Handler & Smith	Biochemistry-clinical application
D. Voet and J C Voet	Biochemistry

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FIRST SEMESTER (July 2018 – December 2018)

PAPER – III: Microbiology [Credit: 4 and Maximum Marks: 80]

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UNIT-I General characteristics of fungi, classification of fungi, life cycle of selected fungal genus (*Aspergillus*, *Pencillium*, *Fusarium* and *Mucor*). Economic importance of fungi. Fungi and bioremediation, parasitism, mutualism and symbiosis with plants and animals. Heterothallism, sex hormone in fungi, Mycorrhiza, VAM.
Algae: Distribution, classification, reproduction, ecology and importance.

UNIT-II Morphology and ultra structure of bacteria:
Morphological types, cell wall of archaebacteria, gram negative, gram positive eubacteria.
Cell membranes – structure, composition and properties. Structure and function of flagella, cilia, pili, gas vesicles.
Cyanobacteria, protozoa, mycoplasma and Rickettsia.
Gene transfer mechanisms: transformation, transduction, conjugation and transfection.
Plasmids: F- factors, colicins and col factors, plasmids as a vector for gene cloning.

UNIT-III Nutritional types (autotrophs, heterotrophs, phototrophs, chemotrophs), growth curves, measurement of growth, factors affecting growth, generation time, growth kinetics.
Batch and continuous culture, asynchronous, synchronous culture.
Basis of microbial classification, classification and salient feature of bacteria according to Bergey's manual of determinative bacteriology, cyanobacteria, prochlorons and cyanelles.

UNIT-IV Viruses: Structure and classification of viruses; General Concepts: Viral genome, capsids, envelopes, viroids and prions).
 Virus reproductions: Lysogeny and Lytic phase,
 Bacteriophages and their types.
 General Introduction to Plant and animal viruses (TMV, HIV, Hepatitis virus, H1N1 virus, Small Pox virus), hemorrhagic viral fever, Route of transmission of viruses, Laboratory diagnosis and treatment, Antiviral therapy.

Lab Course:

1. Glassware preparation and sterilization techniques- wet heat- dry heat- filter types- laminar flow chamber types- CDC- safety levels.
2. Preparation of liquid & solid media, plating, pouring, inoculation and incubation for growth of microorganism
3. Methods of obtaining pure culture of microorganisms (a) streak plate (b) Pour plate, and (c) spread plate methods
4. Microscopic examination of the microorganisms, identification and staining methods
5. Micrometry and camera lucida drawings
6. Study of bacterial growth by turbidimetry/ spectrophotometry
7. Biomass measurement for fungi
8. Isolation and enumeration of microorganisms from soil by serial dilution agar plating method.
9. Enumeration of viruses by plaque assay technique.
10. Motility of bacteria by hanging drop technique.

Books Recommended:

Microbiology	L.M. Prescott, J.P. Harley and D.A. Klein
General Microbiology	RY Stanier, J L Ingrahamana, ML Wheelis & P. R. Painter
Principles of Microbiology	R.M. Atlas
Microbiology	Peleczar, Chan & Krieg.
General Virology	Luria, Darnell, Baltimore and Campell.
Introduction to Mycology	CJ Alexopoulos and CW Mims
Principles of Virology:	S. J. Flint, V. R. Racaniello, L. W. Enquist, V. R. Rancaniello,
Molecular Biology, Pathogenesis, and Control of Animal Viruses.	. M. Skalka

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FIRST SEMESTER (July 2018 – December 2018)

PAPER – IV: Biology of Immune System [Credit: 4 and Maximum Marks: 80]

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UNIT-I Innate immune mechanism and characteristics of adaptive immune response. Cells of immune system: Hematopoiesis and differentiation, mononuclear cells and granulocytes, antigen presenting cells.
 Primary and Secondary lymphoid organs and tissues.
 Ontogeny and phylogeny of lymphocytes. Lymphocyte traffic.

UNIT-II Antigen receptor molecules: B-cell receptor complex, Immunoglobulin- structure, types and function. T-cell receptor complex. Major Histocompatibility Complex- types, structural organization, function and distribution. Transplantation and Rejection. Complement system.

UNIT-III Antigens: nature of antigens, factor affecting immunogenicity, Haptens and super antigens. Antigenic determinants. Recognition of antigens by T and B cell. Antigen processing. Role of MHC molecules in antigen presentation and co-stimulatory signals. Antigen and antibody interaction.

UNIT-IV Cell mediated immune response. Cytokines and interleukins- structure and function. Immunity to infections. Hypersensitive reactions and their types. Immunodeficiency disorders. Autoimmunity

Lab Course:

1. Identification of cells of immune system
2. Separation of mononuclear cells by Ficoll-Hypaque
3. Identification of Lymphocytes and their subsets
4. Lymphoid organs and their microscopic organization
5. Isolation and purification of Antigens
6. Purification of IgG from serum
7. Estimation of Levels of gamma globulins and A/G ratio in blood
8. Antigen antibody interaction

Books Recommended:

Kuby's Immunology	R.A. Goldsby, T. J Kindt and B. A. Osborne
Immunology- A short Course	E. Benjamini, R. Coico and G. Sunshine
Immunology	Roitt, Brostoff and Male
Fundamentals of Immunology	William Paul
Immunology	Tizard
Immunology	Abbas et al

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SECOND SEMESTER (January 2019 – June 2019)

PAPER – I: Genetics and Molecular Biology [Credit: 4 and Maximum Marks: 80]

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UNIT- I Gene mapping methods: Linkage maps, tetrad analysis, mapping with molecular markers, mapping by using somatic cell hybrids, development of mapping population in plants, complementation analysis. Mutation: Types, mutagens and detection.

Mutant types – lethal, conditional, biochemical, loss of function, gain-of-function, germinal verses somatic mutants, insertional mutagenesis.

UNIT-II DNA replication in eukaryotes and prokaryotes : enzymes involved, replication origin and replication fork, fidelity of replication, extrachromosomal replicons. DNA damage and repair mechanisms: Repair of Base-excision, Nucleotide excisions, Mismatch and Double Strand. p_{53} and p_{21} .

UNIT-III RNA synthesis and processing: enzymes involved, formation of initiation complex, transcription activator and repressor, elongation, and termination, RNA processing, capping, RNA editing, splicing, and polyadenylation, RNA transport.

UNIT-IV Protein synthesis and processing: Ribosome, formation of initiation complex, initiation factors, elongation and elongation factors and their regulation, termination.

Aminoacylation of tRNA, tRNA-identity, aminoacyl tRNA synthetase, and translational proof-reading, translational inhibitors.

Post Translational modification of proteins.

Lab Course:

1. Isolation, purification and estimation of RNA
2. Isolation, purification and estimation of DNA
3. Determination of T_m of nucleic acid
4. Fraction of poly (A) RNA
5. Restriction Mapping
6. Restriction Digestion
7. Ligation
8. DNA molecular size determination

Books Recommended:

Molecular Cell Biology	H. Lodish, A. Berk, SL Zipursky, P. Matsudaira, D. Baltimore, and James Darnell.
Essential Cell Biology	B. Alberts, D. Bray, K. Hopkin and A. Johnson
Molecular Biology of the Cell	B. Alberts, A. Johnson, J. Lewis and M. Raff
Cell and Molecular Biology : Concepts and experiments	Gerald Karp
Molecular Biology of the Gene	JD Watson et al.
Molecular Biology of the Cell The Problems	John Wilson, Tim Hunt
Molecular Biology of the Cell	Bruce Albert's, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, Peter Walter
Genes VIII	Benjamin Lewin

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SECOND SEMESTER (January 2019 – June 2019)

PAPER – II: Bioenergetics & Metabolism [Credit: 4 and Maximum Marks: 80]

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of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I First and second laws of thermodynamics. Gibbs free energy G , free energy change ΔG , endergonic & exergonic reactions. Standard state free energy changes- ΔG , ΔG^0 and $\Delta G'^0$, Relationship between equilibrium constant and $\Delta G'^0$, Feasibility of reactions. ATP- Structure, properties and energy currency of the cell, Importance of Coupled reactions, other high energy compounds.

UNIT-II Carbohydrate metabolism: Glycolysis, Kreb's cycle, glycogenolysis, glycogenesis, pentose phosphate pathway, gluconeogenesis, and glyoxylate pathway. Regulation of carbohydrate metabolism.

UNIT-III Electron transport and oxidation phosphorylation: electron carriers, complexes I to IV, substrate level phosphorylation, mechanism of oxidative phosphorylation. Shuttle system for entry of electron.
Biosynthesis and degradation of Lipids. Regulation of lipid metabolism

UNIT-IV Nitrogen Assimilation: Overview of Nitrogen in biosphere and uptake by organism. Biosynthesis and degradation of amino acids. Regulation of amino acid metabolism
Biosynthesis and degradation of purine and pyrimidine nucleotides.

Lab Course:

1. Protein estimation by Lowry, Bradford and Spectrophotometric method
2. Estimation blood cholesterol
3. Estimation of sugar by Nelson- Somogy and Benedict's reagent
4. Isolation and estimation of lipid from seeds and egg.
5. Estimation of inorganic and total phosphorus by Fiske-Subba Rao method
6. Assay of phosphatases in blood and seeds
7. Urease estimation in plant tissues

Books Recommended:

Principles of Biochemistry	Nelson, Cox and Lehninger
Biochemistry	G. Zubay
Biochemistry	Stryer
Biochemistry	Garrett and Grosham
Text book of biochemistry	West, Tood, Mason & Bbruglen
Biochemistry	White,Handler & Smith
Biochemistry with clinical application	D. Voet and J C Voet
Enzymes	Dixon and Webb
Fundamentals of Enzymology	Price and Steven
Practical biochemistry	Plummer
Enzyme biotechnology	G. Tripathi
Enzyme Reaction Mechanism	Walsh
Enzyme catalysis and regulation	Hammes

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SECOND SEMESTER (January 2019 – June 2019)

PAPER- III: Instrumentation and Molecular Techniques

[Credit: 4 and Maximum Marks: 80]

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- UNIT-I** Centrifugation: Principle, techniques. Preparative, analytical and ultracentrifuges, sedimentation coefficient and factors affecting sedimentation coefficient. Application of centrifugation.
Photometry: Basic principles of colorimetry, UV- visible spectrophotometry & IR-spectrophotometry. Spectrofluometry
Atomic absorption spectroscopy: Principle, Instrumentation and applications
- UNIT-II** Microscopic techniques: light microscopy, phase-contrast microscopy, scanning and transmission electron microscopy, different fixation and staining techniques for EM, freeze-etch and freeze-fracture methods for EM, image processing methods in microscopy.
- UNIT-III** Chromatography: Paper and Thin Layer Chromatography. Gel filtration, Ion exchange chromatography and Affinity chromatography. Gas-liquid chromatography and HPLC. Histochemical and immunotechniques: Detection of molecules using ELISA, RIA, western blot, immunoprecipitation, flow cytometry and immunofluorescence microscopy, detection of molecules in living cells, *In situ* localization; FISH and GISH.
- UNIT-IV** Electrophoresis: Agarose, PAGE, 2D-E.
Radioactivity: GM counter, liquid Scintillation counter, solid Scintillation counter, gamma counters.
Lyophilization: Principle, instrumentation and applications.
Microtomy: types, principle and applications

Lab Course:

- Verification of Beers Law
- Determination of absorption maxima
- Quantitative determination, Enzyme kinetics
- Amino acid and carbohydrate separation by paper and TLC
- Ion exchange and gel filtration chromatography
- SDS Polyacralamide Gel Electrophoresis
- DNA electrophoresis
- Isoenzymes
- Separation of sub-cellular organelles by differential centrifugation.
- Isolation of DNA and Agarose gel Electrophoresis
- Amplification of RAPD and AFLP markers.

- Isolation of RNA and Electrophoresis of RNA on denaturing gels.
- cDNA synthesis and cloning
- Isolation of Protein and SDS-PAGE
- In vitro DNA ligation, transformation of E. coli
- Characterization of transformants: DNA gel electrophoresis, Restriction map analysis

Books Recommended:

- | | |
|---|--|
| • K Wilson and John Walker | Practical Biochemistry: Principles & Techniques |
| • RF Boyer | Biochemistry Laboratory: Modern Theory & Techniques |
| • S Carson, H Miller and D Scott | Molecular Biology Techniques: A Classroom Laboratory Manual |
| • TC Ford and J. M. Graham | An Introduction to Centrifugation |
| • R Baserga and D Malamud | Autoradiography: techniques and application |
| • T Chard | An Introduction to Radioimmunoassay and Related Techniques , Volume 6 |
| • MD Bruch | NMR Spectroscopy Techniques |
| • BA Wallace and R William | Modern Techniques for Circular Dichroism and Synchrotron Radiation, Volume 1 |
| • J Sambrook, EF Ritsch and I Maniatis | Molecular cloning: A Laboratory Manual |
| • PD Dabre | Introduction to Practical Molecular Biology |
| • JD Watson, NH Hopkins, JW Roberts, JA Steitz and AM Weiner | Molecular Biology of Gene (4 th Edition) |
| • J Darnell, H Lodish and D Baltimore | Molecular Cell Biology (2 nd Edition) |
| • B Alberts, D Bray, J Lewis, M Raff, K Roberts and JD Watson | Molecular Biology of the Cell (2 nd Edition) |
| • Benjamin Lewin | Gene VII |
| • JM Walker and R Rapley | Molecular Biology and Biotechnology |
| • SB Primrose | Molecular Biotechnology |

M. Sc. Biochemistry

SECOND SEMESTER (January 2019 – June 2019)

PAPER- IV: BIOMETRY, COMPUTER AND SCIENTOMETRY

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

Unit-I Introduction to biostatistics. Types of biological data: data on different scales. Frequency distributions. Cumulative frequency distributions. Random sampling. Parameters and statistics. Measures of central tendency and dispersion: Mean, Median, Mode, Range, Variance and Standard deviation. Coefficient of variation. The effects of coding data. Data transformations: Log-transformation, Square-root transformation and Arcsine transformation. Distribution: normal & binomial. Probability: Basic laws of probability, addition law, multiplication law. Probability and frequency.

Unit-II Statistical errors in hypothesis testing. Testing goodness of fit: Chi-square goodness of fit. Heterogeneity Chi-square. The 2 x 2 contingency table. One sample hypothesis. Two-sample hypothesis. Testing for difference between two means (t-test). Testing for difference between two variances (F-test). The paired sample t-test. Multiple-sample hypothesis (ANOVA): Single factor and two factors ANOVA. Multiple comparisons: Duncan's multiple-range tests. Simple linear regression. Regression vs. Correlation. Regression equation. Interpretations of regression functions. Simple linear correlation. The correlation coefficient.

Unit-III Introduction to MS-Office software: Word processing; creating new document, editing documents, adding graphics to documents, Word tables. Management of Workbook & Worksheets; Applications, Features, Using formulas and functions, Features for Statistical data analysis, Generating charts/ graph. Presentation software; Working in PowerPoint, Creating new presentation, working with slides.

Unit-IV Introduction to Internet and Applications. Basics of internet, e-mailing, Search engine – Google and Yahoo; Pub med, Scopus, Web of Science, Google Scholar, Indian Citation Index, Science Citation Index (SCI), h-index, i-10-index. Journal Impact Factor (JIF). Introduction to Plagiarism and Cyber laws.

Lab Course:

1. Exercises for data distribution
2. Exercises for computation of measures of central tendency
3. Exercises for computation of measures of variability
4. Computation of correlation coefficient, r , and regression constants
5. Data analysis by ANOVA and multiple-range tests
6. Hypothesis testing by t-test, F-test, and Chi-square test
7. Graphical presentation of data using a suitable package
8. Statistical analysis of a data using a suitable package
9. Preparation of document using a suitable package
10. Preparation of slides using a suitable package

Books Recommended:

Campbell RC	Statistics for biologists
Zar JH	Biostatistical Analysis
Wardlaw AC	Practical Statistics for Experimental Biologists
Snedecor GW & Cochran WG	Statistical Methods
Sokal RR & Rohlf FJ	Introduction to Biostatistics
Sumner M	Computers: Concepts & Uses
White R	How Computers Work
Cassel P et al.	Inside Microsoft Office Professional
Coleman P and Dyson P	Mastering Internets
Gralla P	How the Internet Works
Shelly GB, Vermaat ME, Cashman TJ	Microsoft 2007: Introductory Concepts & Techniques
Habraken J	Microsoft Office 2003 All in One
	Microsoft Office 2010 In Depth
Gilmore B	Plagiarism: Why it happens, How to prevent it?
Buranen L & Roy AM	Perspectives on Plagiarism & Intellectual Property in a Post-Modern World
Kumar Anupa P	Cyber Law
Sood V	Cyber Law Simplified

M. Sc. Biochemistry
THIRD SEMESTER (July 2019 – December 2019)
PAPER – I: Genetic Engineering [Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- UNIT- I** Molecular tools and their applications: restriction enzymes, modification enzymes.
Molecular techniques: gel electrophoresis, polymerase chain reaction, DNA sequencing, DNA microarray.
- UNIT-II** Gene cloning vectors: plasmids, lambda phage as vector, M13 phage as vector, cosmids, artificial chromosomes (BAC, PAC, YAC).
Library:
1. Genomic library: genomic DNA library construction and screening methods.
2. cDNA library: cDNA library construction and screening.
Study of gene regulation: reporter assays.
Expression strategies for heterologous genes: vector engineering and codon optimization, host engineering, in vitro transcription and translation.
- UNIT-III** Processing of recombinant proteins: recombinant proteins purification, refolding, characterization and stabilization
Site directed mutagenesis, protein engineering, Gene knockout technique
- UNIT-IV** 1st generation- RFLP, RAPD, 2nd generation- AFLP, SSR, STS (Sequence Tagged Strands), microsatellites, SCAR (Sequence characterized amplified regions), 3rd generation- SNP, SSCP (Single strand conformational polymorphism), 4th generation- DArT.
Hybridization techniques: western blot, northern blot, southern blot.

Lab Course:

1. Bacterial culture and antibiotic selection media. Preparation of competent cells
2. Isolation of plasmid DNA.
3. Isolation of Lambda phage DNA.
4. Quantitation of nucleic acids.
5. Agarose gel electrophoresis and restriction mapping of DNA.
6. Construction of restriction map of plasmid DNA.
7. Cloning in plasmid/phagemid vectors.
8. Isolation of RNA.
9. Synthesis of cDNA.
10. RAPD analysis by PCR.

Books Recommended:

Genes VIII	Benjamin Lewin
An Introduction to Genetic Engineering	DST Nicholl
Principles of Gene Manipulation and Genomics	SB Primrose and Richard
Gene Cloning and Manipulation	CJ Howe
Genetic Engineering (Genetics and Evolution)	R Hodge
Introduction to Biotechnology & Genetic Engineering	AJ Nair
Genetic Engineering	A Kumar & N Garg
Biotechnology & Genetic Engineering	L Yount
DNA Microarrays & Gene Expression: from Experiments to Data Analysis and Modeling	P Baldi & G Wesley
DNA Sequencing (Intro. to Biotechniques)	L Alphey
Plant transformation Technologies	CN Stewart, A Touraev, V Citovsky & T Tzfira
Application of Plant Biotechnology: In vitro Propagation, Plant Transformation and Secondary Metabolite Production	A Kumar and SK Sopory
Genetic Transformation of Plants	JF Jackson & HF Linskens
Transgenic Plants: Methods & Protocols	L Pena

M. Sc. Biochemistry

THIRD SEMESTER (July 2019 – December 2019)

PAPER- II: Plant Physiology and Biochemistry [Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- UNIT- I** Sensory photobiology - Structure, function and mechanisms of action of phytochromes, cryptochromes and phototropins; stomatal movement; photoperiodism and biological clocks.
Organization of shoot and root apical meristem; shoot and root development
- UNIT-II** Photosynthesis: Light absorption and energy conversion, photosystems I and II, ATP synthesis, Assimilation of carbon in C₃, C₄ and CAM pathways, Photorespiration
- UNIT-III** Phytohormones: Structure, biosynthesis, molecular mechanisms of Auxin, Gibberellins, Cytokinin, Absciscic acid and Ethylene, Brassinosteroids.
- UNIT-IV** Senescence and Programmed cell death: Senescence; Metabolism and regulation of pigment and nucleic acid, PGR regulation, SAG. PCD; Formation of TE and mobilization of cereal endosperm, Formation of aerenchyma. Signal transduction and PCD

Lab Course:

1. Spectrophotometric determination of chlorophyll-a, chlorophyll-b and total chlorophyll in young, mature and senescent leaves.
2. Kinetin estimation by cucumber cotyledons expansion bioassay.
3. Auxin bioassay using wheat coleoptiles.
4. GA bioassay by inducing *de-novo* synthesis of Amylase in de-embryonated seeds of wheat.
5. Estimation of mono, di and total phenols in the young and aged leaves.

6. Estimation of Guaiacol peroxidase activity in fresh and aged seeds.
7. Determination of Superoxide dismutase levels in the healthy and deteriorated seeds.
8. Estimation of metal toxicity induced changes in the AOS levels in leaf tissues.
9. Determination of Nitrate reductase activity in leaf tissues.
10. Separation of isozymes of SOD and GPX.

Books Recommended:

Fosket DF	Plant Growth & Development
Foyer CH	Photosynthesis
Bacon KE	Photosynthesis: Photobiochem. & Photobiophysics
Leopold AC & Kriedemann PE	Plant Growth & Development
Moore TC	Biochemistry & Physiology of Hormones
L Taiz & E Zeiger	Plant Physiology
BB Buchanan, W Gruissem & RL Jones	Biochemistry and Molecular Biology of Plants
MB Wilkins	Advanced Plant Physiology
JA Hopkins	Introduction to Plant Physiology
FB Salisbury & CW Ross	Plant Physiology
Hans-Walter Heldt	Plant Biochemistry & Molecular Biology

M. Sc. Biochemistry
THIRD SEMESTER (July 2019 – December 2019)

PAPER- III: Nutritional and Environmental Biochemistry
[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- UNIT-I** Composition of balanced vegetarian and non-vegetarian diets; recommended dietary allowance (RDA) for different categories of the human beings.
Food preservation standards, food adulterations and precautions, government regulations on preservation and quality of food.
Food processing and loss of nutrients during processing and cooking.
Basal metabolism and methods of measuring basal metabolic rate (BMR); energy requirements during growth, pregnancy, lactation and various physical activities.
- UNIT- II** Nutritional aspects of the carbohydrates, lipids and protein: nutritive value, requirements, and functions.
Nutritional aspects of the vitamins and minerals: requirement and functions
Malnutrition, its implications, relationship with dietary habits and prevention.
Disorders related to the nutrition: Protein energy malnutrition, Starvation, Obesity.
- UNIT- III**
Environmental Pollution: Types, Outdoor and indoor Air pollution, sources, structure and control strategies. Water and Soil Pollution.

Eco-toxicology and its environmental significance.
Xenobiotic metabolism, Phase I reaction – oxidation – reduction, hydrolysis and hydration. Phase II reaction – conjugation and methylation.

UNIT- IV

Pesticide toxicity – insecticides, fungicides, herbicides and biopesticides.
Toxicology of food additives.
Metal toxicity – arsenic, mercury, lead and cadmium.
Toxicity testing – Test control, genetic toxicity testing.
Occupational toxicology: Occupational hazards and their assessment.

Lab Course:

1. Separation and purification of sub-cellular organelles and assay of marker enzymes.
2. Protein fractionation - salt, solvent and isoelectric precipitation.
3. Identification and assay of certain toxicants.
4. Effect of various toxicants on serum enzymes and proteins
5. Effect of various toxicants on liver and kidney metabolism
6. Estimation of carbohydrate, protein and fat in food materials.
7. Titrimetric method of ascorbic acid estimation in fruit.
8. Separation of casein protein from milk

Books Recommended:

LG Corkerhem and BSS Shane	Basic Environmental Toxicology
T Shibamoto & L F Bzeidan	Introduction to Food Technology
M. Stipanuk	Biochemical, Phys. & Mol. Aspects of Human Nutrition
Tom Brody	Nutritional Biochemistry
DA Bender	Nutritional Biochemistry of the Vitamins
R.L. Pike and M.L. Brown	Nutrition: An integrated approach -
G.P. Talwar	Text book of Biochemistry and Human Biology
DWS Wong	Mechanism and theory in food chemistry
M.S. Banji N P. Rao & V. Reddy	Text book of Human Nutrition
Linten	Nutritional Biochemistry and Metabolism

M. Sc. Biochemistry

THIRD SEMESTER (July 2019 – December 2019)

PAPER - IV: Enzymology [Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I General properties and effects of pH, substrate and temperature on enzyme catalyzed reactions.

Kinetics of catalyzed reaction: Single substrate reactions, bisubstrate reactions, concept of Michaelis - Menten, Briggs Haldane relationship, Determination and significance of kinetic constants, Limitations of Michaelis-Menten Kinetics,

Concept of convergent and divergent evolution of enzyme.

UNIT-II Enzyme Turnover and methods employed to measure turnover of enzymes, significance of enzyme turnover.

Protein – ligand binding, analysis of binding isotherms, cooperativity phenomenon, Hill and Scatchard plots.

Multienzyme system : occurrence, their properties , mechanism of action & regulation;

Pyruvate dehydrogenase complex, fatty acid synthetase complexes.

Mechanism of action of lysozyme, chymotrypsin, carboxypeptidase and DNA polymerase

UNIT-III General mechanisms of enzyme regulation

Allosteric enzymes, sigmoidal kinetics and their physiological significance, symmetrical and sequential modes for action of allosteric enzymes and their significance.

Water soluble enzymes and their coenzymes. Metallo enzymes.

Immobilized enzymes and their industrial applications.

Enzyme modeling; WHATIF, Verify3d, PROSA and DOPE score

UNIT-IV Enzymes of Industrial Importance; their source, characteristic properties, functions and uses.

Enzymes used in leather, paper, textile industries.

Enzymes in baking, brewing, Alcohol products; enzymes in detergents, starch and animal feeds.

Amylases, cellulases, catalase, pectinase, lipase, protease, xylanase, laccase, beta glucanase

Lab Course:

1. Estimation of enzymes
2. Separation, purification of sub-cellular organelles & assay of marker enzymes.
3. Methods of purification of an enzyme - ion-exchange, gel filtration
4. Test of homogeneity by SDS-PAGE
5. Kinetics of an enzymatic reaction
6. Effect of various toxicants on serum enzymes and proteins
7. Enzyme modeling: Validation Criteria by WHATIF, Verify3d, PROSA and DOPE score
8. Verification of Ramachandran Plot: Estimation of interaction energy per residue by PROSA and Verify3D.
9. Enzyme packing quality: Assessed by WHATIF.

Books Recommended:

Brandon and Tooze	Introduction to Protein Structure
Campbell	Discovering Genomics, Proteomics and Bioinformatics,
Dan Gusfield	Algorithms on Strings Trees and Sequences
Lesk, A.M	Introduction to Protein Architecture
Mcpherson, A.	Introduction of Molecular Crystallography
Pennington	Proteomics from Protein Sequence to Function
Durbin, Eddy, Anders & Graeme	Biological Seq. Analysis: Probabilistic Models of Proteins & Nucleic Acids
S.A. Bbernhard	The structure and function of enzymes

J. Palmer	Enzymes: biochemistry, Biotechnology, Clinical chemistry
M Dixon, EC Webb, CJR Thorne & KF Tipton	Enzymes
Alan Fersht	Enzyme structure and Mechanism
Christopher Walsh	Enzymatic reaction mechanism
Eisenthal and Danson	Enzyme Assay: A Practical Approach
M. Stipanuk	Biochemical, Phys. & Mol. Aspects of Human Nutrition
G.P. Talwar	Text book of Biochemistry and Human Biology

M. Sc. Biochemistry

FOURTH SEMESTER (January 2020 – June 2020)

PAPER – I: Plant Biotechnology [Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT- I Introduction to cell and tissue culture: Tissue culture media (composition and preparation), explant preparation, Callus and suspension culture, cytodifferentiation and organogenic differentiation, somatic embryogenesis, micropropagation.
Shoot tip culture: Rapid clonal propagation and production of virus free plant.

UNIT-II Embryo culture and embryo rescue.
Anther, pollen and ovary culture for production of haploid plants & homozygous lines.
Protoplast isolation, culture and fusion; selection of hybrid cells and regeneration of hybrid plants; symmetric and asymmetric hybrids, cybrids.
Germplasm conservation: Cryopreservation & slow growth cultures.
Chloroplast Transformation: Advantages, vectors, success; tobacco & potato.

UNIT-III Plant transformation technology: Plant transformation technology: basis of tumor formation, hairy root, features of Ti and Ri plasmids, mechanism of DNA transfer, role of virulence genes, use Ti and Ri as vectors, binary vectors, use of 35S and other promoters, use of reporter genes, particle bombardment, electroporation, microinjection.

UNIT-IV Applications of plant transformation for productivity and performance: herbicide resistance, insect resistance, long shelf-life of fruits. Bt genes, Non-Bt like protease inhibitors & amylase inhibitors, virus resistance, nucleocapsid gene, disease resistance, PR (Pathogenesis Related) proteins, nematode resistance, abiotic stress, male sterile lines

Lab Course:

1. Preparation of culture media.
2. To perform meristem/ bud culture, shoot multiplication & rooting phenomenon.
3. To study organogenesis.
4. To perform somatic embryogenesis.

5. To study the process of plantlet acclimatization.
6. To perform embryo culture.
7. To study the process of anther culture development.
8. Study of molecular markers.
9. Extraction of DNA from plant cultures.
10. Estimation & separation of DNA: Agarose gel electrophoresis & spectrophotometer.

Books Recommended:

Razdan MK	Introduction to Plant Tissue Culture
Vasil IK	Plant Cell and Tissue Culture
Bhojwani SS and Razdan MK	Plant Tissue Culture
Fu TJ, Singh G and Curtis WR	Plant Cell & Tissue Culture for the production of Food Ingredients
Hammond, McGarvP & Yusibov	Plant Biotechnology
Singh BD	Biotechnology: Expanding Horizons
RH Smith Plant Tissue Culture	Techniques and Experiments
L Kyte and J Kleyon	Plants from Test Tubes: An Introduction to Micropropagation
M Smith	Plant Propagator's Bible
MR Ahuja	Micropropagation of Woody Plants
YPS Bajaj	Trees III
YPS Bajaj	Trees IV

M. Sc. Biochemistry

FOURTH SEMESTER (January 2020 – June 2020)

PAPER- II: Seed Science Technology [Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

- UNIT- I** Seed development: Phases of development, Maturation; accumulation of desiccation related compounds, ABA regulation. Seed Dormancy: Physiological and molecular basis, Testa, Endosperm, Aleurone layers & Hormonal cross talk in dormancy. Alleviation of dormancy; Protein oxidation. Dormancy breaking chemicals and mechanism.
- UNIT-II** Seed Germination: Pre-germination, Germination and post germination Metabolism. Reactivation of the metabolic pathway. Cellular repair. Hormonal regulation and metabolism; GA & ABA, ROS metabolism.
- UNIT-III** Seed Ageing: Seed storage physiology: Orthodox & Recalcitrant; ROS metabolism, Mechanism of desiccation tolerance, dehydrins/LEA/peroxiredoxin, HSPs, Sugars. Longevity markers; β - mercaptopyruvate sulfurtransferase (MST), L-isoaspartyl O-methyltransferase (PIMT).
- UNIT-IV** Seed Technology: Priming technology; biochemical and molecular aspects. Cryobanks, Cryopreservation of seed and embryo; Cryoprotective molecules, Vitrification, Encapsulation and Drying. Synthetic seeds.

Lab Course:

1. Osmotic and chemical priming effect on seed germination.
2. To observe inhibition of root growth in dose dependent salt stress and its comparison with the control.
3. Testing seed viability and vigour by:
 - (a) Germination
 - (b) Triphenyl tetrazolium test
 - (c) Specific conductance of leachates and
 - (d) Germination Index
4. Lipid peroxidation in ageing seeds.
5. Staining of superoxide and hydrogen peroxide in roots of salt stressed and control seeds.
6. Quantitative and qualitative estimation of antioxidant enzymes in seeds:
 - (a) SOD
 - (b) Guaiacol peroxidase and
7. Peroxidase assay by tissue printing method.

Books Recommended:

J.D. Bewley & M. Black	Physiology & Biochemistry of Seeds
J.D. Bewley & M. Black	Seeds: Physiology of Development & Germination
Black et al.	Desiccation and Survival of Plants: Dying without Drying
P.K. Agrawal & M. Dadlani	Techniques in Seed Science & Technology
FAO Report 113	Ex-situ storage of seeds, pollen & in-vitro cultures
Copeland & McDonald	Seed Science & Technology
R.L. Agrawal	Seed Technology
J. Kigel & G. Galili	Seed Development & Germination
W. Ayad et al.	Molecular Genetic Techniques for Plant Genetic resources
E.E. Benson	Plant Conservation Biotechnology
D. E. Fosket	Plant Growth & Development
R.B. Taylorson	Recent Adv. in the Development & Germination of Seeds
McDonald & Copeland	Seed Technology Laboratory Manual
Khullar & Thapliyal, R.C.	Forest Seed
L. Schmidt	Guide to Handling of Tropical & Sub-tropical Forest Seed

M. Sc. Biochemistry

FOURTH SEMESTER (January 2020 – June 2020)

Special Paper PAPER - III (A): Clinical Biochemistry and Endocrinology

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT-I Plasma proteins – Properties, functions and their variations in diseases, Plasma lipids and lipoproteins, Interrelationship of lipids, lipoproteins and apolipoproteins. Erythropoiesis, abnormalities in blood formation. Anemias. Hemoglobinopathies. Cerebrospinal fluid – composition in health and diseases. Clinical enzymology - Plasma enzymes in diagnosis and prognosis, Isoenzymes in health and diseases (Liver, cardiac and skeletal muscle enzymes)

UNIT-II Liver function tests, their significance, Liver diseases – Jaundice, hepatitis, gall stones, cirrhosis and fatty liver. Free radical mechanism and role of reactive oxygen species in diseases. Role of liver in metabolic regulation and drug metabolism. Clinical chemistry of new born.

Kidney – Renal hormones –Renin, erythropoietin and angiotensin. Investigations of renal functions, biochemical investigation of renal disorders. Nephritis, nephrotic syndrome and urolithiasis. Compensatory mechanism for acidosis and alkalosis.

Gastrointestinal hormones - Gastrin, secretin and cholecystokinin. Disorders of gastric function, methods of evaluation. Pancreatic exocrine secretions, pancreatic diseases, steatorrhea. Malabsorption syndrome – tests for their evaluation and significance.

UNIT-III Pancreatic hormones – Biosynthesis of insulin, regulation of secretion of insulin and glucagon, their role in carbohydrate, lipid and protein metabolism. Endocrine disorders of pancreas – Diabetes mellitus, melliturias, hypoglycemia. Glucose tolerance test.

Thyroidal hormones – Chemistry, function and metabolism. Hypo and hyper thyroidism, tests for thyroid function. Parathyroid hormones – Parathormone and calcitonin, their role in calcium and phosphate metabolism, abnormalities of parathyroid functions and methods of evaluation.

Adrenals - Chemistry and biosynthesis of adrenal medullary and adrenal cortical hormones. Disorders of adrenal cortex and adrenal medulla, tests for the evaluation of adrenal functions. Biochemical effects of tumours.

UNIT-IV Synthesis, secretion, transport and biological actions of hypothalamic, adeno-hypophyseal and neurohypophyseal hormones. Hypothalamic disorders. Pituitary - Clinical syndromes and their evaluation. Pituitary hormones – Melatonin and serotonin. Chemistry, biosynthesis and role of androgens, estrogens and progesterone. Hormonal regulation of menstrual cycle, Hormonal contraception. Placental hormones. Biochemistry of reproductive disorders, pregnancy toxemia, pregnancy tests.

Lab Course:

1. Assay of Alkaline and Acid Phosphates
2. Estimation of blood glucose by GOD and POD method
3. Various types of glucose tolerance tests.
4. Estimation of SGOT, SGPT, LDH and CPK, Serum Amylase enzymes
5. Estimation of HDL- cholesterol, LDL- cholesterol.
6. Estimation of uric acid and creatinine in plasma.
7. Estimation of urine and blood bilirubin.
8. Effect of various toxicants on serum enzymes and proteins
9. Effect of various toxicants on liver and kidney metabolism
10. Purification of protein hormones
11. Assay of steroid dehydrogenase
12. Isolation and characterization of steroid
13. Sperm count
14. Demonstration of estrus cycle study by vaginal smear technique
15. Histological / Histochemical / Cytological study of Endocrine gland

Books Recommended:

Experimental Endocrinology: Zarrow, M.X; Yochin, J.M and Machrth, J.I

Essential techniques in reproductive physiology and Endocrinology: Chinoy, N.J, Rao, M.V, Desarai, K.J and High land, H.N
 Biochemistry: L. Stryer
 Textbook of Biochemistry with Clinical Correlations: T.M. Devlin
 Lippincott's Illustrated Reviews in Biochemistry: P.C.Champe, R.A.Harvey and D.R.Ferrier
 Harper's Biochemistry: R.K.Murray, D.K.Granner, P.A. Mayes and V.W.Rodwell.
 Clinical Laboratory Science Review: Robert R. Harr
 Fundamentals of Clinical Chemistry: C.A. Burtis, E.R. Ashwood Tietz
 Notes on Clinical Chemistry- Principles of Internal Medicines: Whitby, Smith, Beckett, Walker, Harrison
 The structure and function of enzymes: S.A. Bbernhard
 Enzymes- biochemistry, Biotechnology, Clinical chemistry: J. Palmer
 Enzymes: Dixon, Webb, Thorne & Tipton
 Enzyme structure and Mechanism: Alan Fersht
 Enzymatic reaction mechanism: C. Walsh, F. Pub
 Basic Environmental Toxicology: Basic Environmental Toxicology: L. G Corkerhem and B.SS Shane
 Introduction to Food Technology: T. Shibamoto & L F Bzeidan
 Enzyme Assay: A Practical Approach: Eisenthal and Danson
 Biochemical, Physiological & Molecular Aspects of Human Nutrition: M. Stipa

M. Sc. Biochemistry
FOURTH SEMESTER (January 2020 – June 2020)
Special Paper: PAPER- III (B): Nutraceutical Biochemistry and Functional Foods
[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

Unit-I: Introduction to Nutraceuticals as Science:

Historical perspective, classification, scope and future prospects. Scrutinising the term 'nutraceutical', Regulation of various countries. Medicinal Plants: Ethnomedicine in India, Applied aspects of the Nutraceutical Science. Sources of Nutraceuticals. Relation of Nutraceutical Science with other Sciences: Medicine, Human physiology, genetics, food technology, chemistry and nutrition

Unit-II: Properties, structure and functions of various Nutraceuticals:

Glucosamine, Octacosanol, Lycopene, Flavonoids, Carnitine, Melatonin and Ornithine alpha, ketoglutarate. Use of proanthocyanidins, grape products, flaxseed oil as Nutraceuticals.
 Nutraceutical Industry and Market Information, New technologies in development of Nutraceuticals and functional foods
 Functional Foods, Scope of Genetic engineering, Nutritional Genomics

Unit-III: Food as remedies

Nutraceuticals bridging the gap between food and drug, Special Dietary Needs, Disease and Nutrition; Nutraceuticals in treatment for cognitive decline, Nutraceutical remedies for common disorders like Arthritis, Bronchitis, circulatory problems, hypoglycemia, Nephrological disorders, Liver disorders, Osteoporosis, Psoriasis and Ulcers etc. Brief idea about some Nutraceutical rich supplements e.g. Bee pollen, Caffeine, Green tea, Lecithin, Mushroom extract, Chlorophyll, Kelp and Spirulina etc.

Unit-IV: Anti-nutritional Factors present in Foods

Types of inhibitors present in various foods and how they can be inactivated. General idea about role of Probiotics and Prebiotics as nutraceuticals. Recent advances in techniques & feeding of substrates. Assessment of nutritional status and Recommended Daily allowances.

Lab Course:

- Identification using characteristic features of nutraceutically important plants like; *Phyllanthus emblica*, *Curcuma longa*, *Zinziber officinalis*, Solanaceae (*Withania somnifera*), *Aloe vera*, Liliaceae (*Alium sativum*), Lamiaceae (*Ocimum sanctum*), Apiaceae (*Coriandrum sps*) and Liliaceae (*Asparagus sps.*), *Centella asiatica*.
- Study of following Parasites/ Vectors/ pests: Identification, Habits and control measures (museum Specimens / slides): *Entamoeba histolytica*, *Taenia sps*, *Ascaris lumbricoides*, *Ancylostoma dueodenaei*, *Trichinella spiralis*, *Trichura trichuris*, Mosquito (*Culex and Anopheles*), House fly, Green bottle fly, Head Louse, Cockroach (*Periplanata & Blatta*), bed bug, *Mus sps.* (Mouse) and *Rattus sps.* (House rat)
- Reactions of mono, di and polysaccharides and their identification in unknown mixtures
- Determination of Acid value, Saponification and Iodine number of natural fats & oils.
- Estimation of proteins with Bradford's and other methods.
- Extraction and estimation of total sugars from food products (dairy product, fruit juices, bread).
- TLC separation of Plant pigments – Curcumin and carotene.
- To isolate DNA and RNA from given plant/ animal material and estimate DNA by Diphenylamine (DPA) method and RNA by Orcinol reagent
- Extraction, purification and evaluation of activity of any one digestive enzyme (e.g. Beta amylase from sweet potato)
- Estimation of ascorbic acid from lemon & amla juice by titration method
- Estimation of crude fat contents of foods by Soxhlet's method (Butter, Margarine, edible oil).
- Estimation of total Nitrogen of foods by Kjeldahl and Micro Kjeldahl methods.
- Chromatography: Paper, TLC, adsorption, ion exchange, gel filtration, affinity, GC & HPLC.
- Separation of Milk proteins on Native and SDS gels.
- Preparation of plasmid DNA from given sample and its digestion by restriction enzymes and separation of DNA fragments by gel electrophoresis

Books Recommended:

- | | |
|------------------------|---------------|
| 01. Stryer E.A., | Biochemistry |
| 02. Zubay, Geoffrey L. | Biochemistry, |

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| 03. Greenberg David M. | Metabolic Pathways, Vol 3 |
| 04. Todd and others, | Clinical Diagnosis and Management, 17th Ed, |
| 05. Gopalan C., et al | Dietary Allowances for Indians, NIH, Hyderabad. |
| 06. Anita F.P. | Clinical Dietetics and Nutrition, 4th Ed, 1997, |
| 07. Devlin, T.M. | Text Book of Biochemistry with Clinical Correlation, |
| 08. Mahan, L.K. & Ecott- Stump, S. [Ed.] | Krause's Food, Nutrition and Diet Therapy |
| 09. Lehninger | Nutrition Concepts & Controversies, |
| 10. Davidson, S. Passmore, & Turswell | Nutrition and dietetics |
| 10. Goodhearth R., S. Shills | Modern Nutrition in health and disease |
| 12. Nelson and Cox, 2000, | Lehninger's Principles of Biochemistry, |
| 13. Robert E.C. Wildman, | Handbook of Nutraceuticals and Functional Foods |
| 16. Rapport and B. Lockwood | Nutraceuticals |
| 15. W. Jeffrey, Hursts | Methods of Analysis for Functional Foods and Nutraceuticals |
| 16. M. Maffei (Ed.) | Dietary Supplements of Plant Origin |
| 17. Gunzler and Williams | Handbook of Analytical Techniques Vol. I,II , |
| 18. Thomson | Herbal Medicines PDR 3rd ed. - |
| 19. Gary, M & Giintert, E. | Active Compounds in Foods Chemistry and Sensory Properties - I |
| 20. Israel Goldberg | Functional foods, designer foods, pharma foods, Nutraceuticals, |
| 21. P.D. Dabre | Introduction to Practical Molecular Biology, |
| 22. Ellyn Daugherty | Biotechnology Science for the new Millennium, |
| 23. T K Attwood, D J PSmith | Bioinformatics Introduction -. |
| 24. Primrose and RM Twyman | Principals of Gene Manipulation and Genomics. |
| 25. Massimo Maffei | Dietary Supplements of Plant origin: a nutrition and health approach |
| 27. CCRUM | Herbal Drugs: Potential Antimalarial Herbal Drugs from South Asia. |

M. Sc. Biochemistry

FOURTH SEMESTER (January 2020 – June 2020)

Special Paper: PAPER- IV (A): Advanced Immunology, diagnostics and prophylaxis

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

UNIT- I Clonal selection theory- concept of antigen specific receptor. Organization and expression of immunoglobulin genes. Generation of antibody diversity. Light and heavy chain gene recombination. Recombination Signal Sequences. Heavy chain constant region genes. Class switching. T-cell receptor diversity.

UNIT-II Membrane and secreted immunoglobulins. Production of polyclonal and monoclonal antibodies- principle, technique and applications. Antibody engineering. Regulation of

immune response by antigen, antibody, immune complex, MHC and cytokines. Immune response to infectious diseases: viral, bacterial and protozoal. Cancer and immune system. Nutrition and Immune response.

UNIT-III Principles of Immunodiagnosis. Antigen-antibody interactions. Precipitation reactions. Haemagglutination. Complement fixation test. Immunofluorescence assay: Fluorescence activated cell sorter (FACS) technique. Radio Immuno and Enzyme Immuno assays. Immunoblotting. Isolation of pure antibodies. Isolation of leucocyte population on density gradient. Effector cell assays. Plaque forming cell assay, ELISPOT assay, leucocyte migration inhibition technique, cytotoxicity assay.

UNIT-IV Active immunization (immunoprophylaxis): Principles of vaccination. Immunization practices. Passive immunization (immunotherapy). Role of vaccine in prevention of diseases: vaccines against important viral, bacterial, protozoan and parasitic diseases. DNA vaccines; Antiviral, antibacterial agents.

Lab Course:

1. Preparation of Parasite Antigen and analysis by PAGE
2. Immunizations and production of antibody
3. Antigen antibody reaction by Double Diffusion, Counter current and IEP, RID & EIA
4. Western Blot Analysis
5. Immunodiagnosis using commercial kits

Books Recommended:

R.A. Goldsby, T.J Kindt & B. A. Osborne	Kuby's Immunology:
E. Benjamini, R. Coico and G. Sunshine	Immunology-A short Course
Roitt, Brostoff and Male	Immunology
William Paul	Fundamentals of Immunology
Stewart Snell	Immunology, Immunopathology and Immunity
Elgert	Understanding Immune System

M. Sc. Biochemistry

FOURTH SEMESTER (January 2020 – June 2020)

Special Paper: PAPER- IV (B): Bioinformatics

[Credit: 4 and Maximum Marks: 80]

(Each theory paper will have questions divided into four sections, A, B, C & D. Section A will have 20 MCQ of 1 mark each covering whole syllabus. Section B will have 8 very short answer questions, two from each unit, of 2 marks each to be answered in two to three lines. Section C will have 8 questions, two from each unit, of 3 marks each. The question has to be answered in about 75 words. Section D will have 4 questions, one from each unit with internal choice, of 5 marks each. The question has to be answered in about 150 words).

Unit I Introduction to bioinformatics and data generation

Bioinformatics and its relation with molecular biology. Examples of related tools (FASTA, BLAST, BLAT, RASMOL), databases (GENBANK, Pub med, PDB) and software (RASMOL, Ligand Explorer).

Data generation; Generation of large scale molecular biology data. (Through Genome sequencing, Protein sequencing, Gel electrophoresis, NMR Spectroscopy, X-Ray Diffraction, and microarray). Applications of Bioinformatics.

Unit II Biological Database and its Types

Introduction to data types and Source. Population and sample. Classification and Presentation of Data. Quality of data, private and public data sources. General Introduction of Biological Databases; Nucleic acid databases (NCBI, DDBJ, and EMBL). Protein databases (Primary, Composite, and Secondary). Specialized Genome databases: (SGD, TIGR, and ACeDB). Structure databases (CATH, SCOP, and PDB sum)

Unit III Data storage and retrieval and Interoperability

Flat files, relational, object oriented databases and controlled vocabularies. File Format

(Genbank, DDBJ, FASTA, PDB, SwissProt). Introduction to Metadata and search; Indices, Boolean, Fuzzy, Neighboring search. Data exchange and integration. Ontologies, interchange languages and standardization efforts. General Introduction to XML, UMLS, CORBA, PYTHON and OMG/LIFESCIENCE.

Unit IV Gene Expression and Representation of patterns and relationship

General introduction to Gene expression in prokaryotes and eukaryotes, transcription factors binding sites. SNP, EST, STS.

Regular Expression, Hierarchies, and Graphical models (including Markov chain and Bayes notes). Genetic variability and connections to clinical data.

Lab Course:

01. Retrieval of sequences from NCBI, EBI and EMBL databases.
02. Retrieval of sequences from NBRF-PIR, SWISSPROT and P databases.
03. Transition and Translation of sequences.
04. Retrieval of genome from genome databases.
05. Exploring DIP and PPI.
06. Exploring BIND and PIM.
07. Exploring MINT and GRID.
08. Analysis of phylogenetic tree
09. Exploring PDB file.
10. Analysis of active site by pymol

Books Recommended:

BAXEVANIS, AD & OUELLETTE, BFF : Bioinformatics: a practical guide to the analysis of genes and proteins. 2nd Ed.. 2002.

BAXEVANIS, AD, DAVISON, DB, PAGE: Current protocols in bioinformatics. 2004.

RDM & PETSKE, GA

ORENGO, C, JONES, D & : Bioinformatics: genes, proteins and computers. 2003

THORNTON, J

Ingvar Eidhammer, Inge Jonassen, : Protein Bioinformatics. 2003

William R Taylor

HIGGINS, D & TAYLOR, W : Bioinformatics: sequence, structure, and databank. 2000.

David Mount : Bioinformatics: sequence and genome analysis. 2004